

Microbiology User handbook

Contents

1.	Intr	oduction6			
2.	2. Change Details				
3.	3. Laboratory location7				
4.	Cor	ntact Details7			
5.	Оре	ening hours, Clinical advice, and results8			
5	.1	Laboratory Opening Hours8			
5	.2	Clinical advice			
5	.3	Urgent samples9			
5	.4	Testing out of hours9			
5	.5	Requesting Tests9			
5	.6 Re	esults			
5	.7	Telephoned results			
5	.8	Turnaround times10			
5	.9	Measurement of Uncertainty10			
6	San	nple Collection			
6	5.1	Viral and bacterial serology tests11			
6	.2	Optimum time of and conditions for collection11			
6	.3	Health and safety issues pertaining to sample collection11			
7	San	nple Containers12			
7	.1	Supply of sample containers12			
7	.2	Selection of appropriate containers13			
7	.3	Labelling of sample containers14			
8	Rec	juest Forms14			
8	.1	Electronic requesting			
8	.2 M	anual requesting16			
8	.3	Anonymous/uniquely identified samples17			
9	San	nple Acceptance and Rejection Criteria18			
10	т	ransportation of Samples			
1	0.1	Transportation of routine samples to the laboratory18			
Act	ive D	ate: 13/06/2025 Review Date: 13/06/2027 CONTROLLED DOCUMENT; UNCONTROLLED WHEN PRINTED OR COPIED			

Q-Pu	ment Title: Microbiology User Handbook se Index: MIC-POL-15 ion Number: 2.1 Author: Olutoye, Yinka	Salisbury NHS Foundation Trust
	.0.1.1 Pneumatic Chute	
	0.1.2 Portering Services	19
	0.1.3 Courier Services	19
10	2 Transportation of urgent samples	19
10	3 High Risk Samples	19
11.	Sample storage	
12.	Quality Assurance in Microbiology	
12.	1 UKAS accreditation	21
12.	2 External Quality Assurance (EQA)	21
12.	3 IQC (Internal Quality Control)	21
12.	4 IQA (Internal Quality Assurance)	21
12.	5 Acceptance testing	21
12.	6 Audits	21
12.	7 Quality indicators	21
13.	Repertoire of Tests (A-Z)	
13.	1 Bacterial Culture, Fungal Culture and Viral Investigations	22
1	Blood culture	22
	Bordetella pertussis	
(Candida Antifungal Susceptibility testing	23
(Cerebro-Spinal Fluid (CSF) Culture	24
(Corneal Scrape Culture	24
I	aecal Samples for bacterial, viral and parasitology tests	25
l	ungal Culture	26
(Gonococcal Culture (GUM clinic only)	27
(Gynaecological Culture	28
I	nfection Control Screening Tests	28
I	ntravenous (IV) Cannula Culture, e.g., CVP line tip	29
1	Neonatal Screen Culture	
I	Parasitology investigation for Enterobius vermicularis	
I	Pus Culture	31
1	Respiratory culture	31
9	taphylococcus and Streptococcus Reference Service (and PVL)	32
	terile Fluid Culture	
	wabs for Microscopy, Culture and Sensitivities (M, C &S)	
	issue Culture	
	e Date: 13/06/2025 Revie CONTROLLED DOCUMENT; UNCONTROLLED WHEN PRINTED OR C 2 of 82	w Date: 13/06/2027 COPIED

Document Title: Microbiology User Q-Pulse Index: MIC-POL-15	Handbook	NHS Salisbury
Revision Number: 2.1	• •	NHS Foundation Trust
TB Culture (Sputum/ BAL/ Tissi	ue/ Pus)	
Urine Culture		
Urinary Parasitology (Schistosc	omiasis)	
13.2 Antigen and Molecular tests		
16s PCP PCR/ 18s Pan-fungal P	CR	
Acanthamoeba Detection		
Adenovirus PCR		
Chlamydia trachomatis Infectio	on	
Chlamydia trachomatis (LGV)		40
COVID-19 PCR		40
Cryptococcus Antigen		
Detection of Viruses in CSF san	nples	41
Full Respiratory Panel (PCR)		41
Meningococcal PCR		
Monkeypox (Mpox) PCR		
Mycobacterium tuberculosis T	3 Fast Track	
Mycobacterium tuberculosis T	3 T-SPOT	
Mycoplasma genitalium PCR		
Parasitology investigation for E	nterobius vermicularis	
RSV Detection		
Urine Antigen Tests: Pneumoco	occal and Legionella	
13.3 Serology and Confirmatory F	PCR tests	47
Antenatal (booking blood) Sero	ology	
Anti-streptolysin titre (ASO Titi	re)	
Avian Precipitins		
Beta-glucan		
BK Quantitative PCR virus (Ren	al patients)	
BK Quantitative PCR virus (Nor	n-renal patients)	
Bordetella pertussis serology		
Bordetella pertussis PCR		
Brucella serology		50
CMV Avidity		51
CMV IgG and/or CMV IgM		51
CMV PCR		52
Active Date: 13/06/2025		Review Date: 13/06/2027

Document Title: Microbiology User Handbook	NHS
Q-Pulse Index: MIC-POL-15 Revision Number: 2.1 Author: Olutoye, Yinka	Salisbury NHS Foundation Trust
EBV Serology	
EBV PCR	53
Fertility Screening	53
Functional Antibodies (Pneumococcal IgG, Haemophilus IgG, Tetanus IgG)	53
Fungal precipitins	54
Haemophilus influenzae Type B Ab	54
Hepatitis A Serology	55
Hepatitis A PCR	55
Hepatitis B surface Antibody	55
Hepatitis B Core Total Antibody	56
Hepatitis B Surface Antigen	56
Hepatitis B e Antigen and Antibody and Hepatitis B core IgM	57
Hepatitis B DNA Viral load	57
Hepatitis C Antibody	57
Hepatitis C PCR Qualitative	58
Hepatitis C Viral load/Genotype	58
Hepatitis D (Delta agent)	59
Hepatitis E IgM and IgG	59
Herpes Simplex Virus 1 and 2 PCR (Qualitative)	59
HIV 1/2 Ab/Ag	60
HIV 1 Pro-Viral DNA	60
HIV 1 RNA Viral Load	61
HIV 1 Genotypic Resistance Test	61
HIV-2 Viral Load	62
HTLV-1 & HTLV-2 Ab	62
Human Herpes Virus-6 DNA Quantitative PCR	62
Human Herpes Virus-7 DNA Quantitative PCR	63
Human Herpes Virus-8 DNA Quantitative PCR	63
Leptospiral serology IgM/Leptospiral PCR	64
Lyme (Borrelia burgdorferi) IgG and IgM	64
Measles Serology IgM	65
Measles Serology IgG	65
Measles PCR	65
Mumps Serology IgG	66
Mumps Serology IgM	66

Active Date: 13/06/2025

NHS

Document Title: Microbiology User Handbook		
Q-Pulse Index: MIC-POL-15 Revision Number: 2.1	Author: Olutoye, Yinka	NHS Foundation Trust
Mumps PCR		67
Non-indigenous mycoses	(e.g., Histoplasma) serology	67
Parasite disease serology		68
Parvovirus B19 lgG and lg	M Serology	68
Parvovirus B19 PCR		68
Pneumococcal PCR		69
Pneumococcal Serology		69
Rabies Serology		70
Rubella Serology		70
Schistosoma Serology		70
Syphilis Serology		71
Syphilis IgM, RPR, TPPA, a	Iternative EIA and Immunoblots	71
Syphilis PCR		72
Toxoplasma gondii Serolo	gy	72
Tropical Disease Serology	(Regional Travel Screen)	72
Varicella zoster Serology	Chickenpox)	73
Varicella zoster PCR		73
13.4 Antibiotic assays		74
Gentamicin Levels		74
Tobramycin Levels		75
Amikacin Levels		75
Vancomycin Levels		76
Teicoplanin level		76
Other antibiotic level, e.g	., Co-trimoxazole	77
Anti-Fungal drug level		77
13.5 Family Planning		78
Sub-fertility semen analys	is	
Post vasectomy semen ar	alysis	79
14. Point of Care Testing (I	20CT)	
15. Patient Consent Disclo	sure	
	otection of personal information	
	S	
5 1	ct	
	biology service and complaints procedure	
Active Date: 13/06/2025 CONTROLLED DO	Re DCUMENT; UNCONTROLLED WHEN PRINTED O	view Date: 13/06/2027 R COPIED

NHS

Docur Q-Pul Revisi	Salisbury NHS Foundation Trust		
17	Associated Documents		
18	Appendix		
Арр	oendix 1- HSE Safety Noti	се	82

1. Introduction

The Microbiology department at the Salisbury District Hospital NHS Foundation Trust provides an analytical and interpretative service on a wide range of clinical specimens for medical and infection control advice to hospital and community health care services. The laboratory also provides microbiological support to the local Health Protection Teams and United Kingdom Health Security Agency (UKHSA) departments. Specialist and Reference test services are used where necessary.

We process over 220,000 specimens each year, many requiring multiple investigations. Our ability to process requests in a timely fashion relies heavily on receiving correctly completed request forms from our users. Your compliance with the guidelines concerning safety, specimen identification and transport will help us to deliver a safe, efficient, and legally defensible service.

In its pursuit of excellence and as part of its continuous quality improvement programme the Microbiology department participates in all relevant internal and external quality assurance schemes.

The repertoire of tests provided by Microbiology support the Trust in its diagnostic and screening programmes. The laboratory is accredited by the Institute of Biomedical Science (IBMS) for Biomedical Scientist training and Biomedical Scientist Specialist training.

It is anticipated that this handbook will provide the information you require to use our service.

2. Change Details

Any copy printed in the wards or GP surgery or other location outside the laboratories control becomes an uncontrolled document and is not managed under the Microbiology Control procedure. It is the responsibility of the copyholder to ensure that any hard copy in their possession reflects the current version available on the intranet sites.

Date	Version Number	Change	Sections affected
	1.0	New format to Qpulse, updated all sections and split into microbiology only	
20/12/24	2	Updated	All sections
03/06/2025	2.1	Addition of MRSA and Norovirus PCR, change of ref lab for <i>B. pertussis</i>	13



Author: Olutoye, Yinka

03/06/2025	2.1	Removal of Dr Flannagan and addition of Dr. Lam	4

3. Laboratory location

Microbiology department is located on Level 4, SDH North.



Image 1: Map of the Microbiology department within the Hospital

4. Contact Details

Position	Name	Telephone Number
Laboratory Manager	Jo Harris	Ext. 4115
Laboratory Administrator:	Julie Wilson	Ext. 4105
Quality Manager:	Yinka Olutoye	Ext. 4104
Consultant Microbiologist Laboratory Director/Deputy Clinical Lead CFSF/ IDPS Clinical Advisor	Dr Julian Hemming	Ext. 4110 (01722 429105)
Consultant Microbiologist/	Dr Paul Russell	Ext. 4101 (01722 429105)

Active Date: 13/06/2025

Document Title: Microbiology User Handbook Q-Pulse Index: MIC-POL-15 Revision Number: 2.1 Author: Olutove, Yinka



Author: Olutoye, finka	
Dr Layth Alsaffar	Ext. 4102 (01722 429105)
Dr. Bosco Lam	Ext. 4102 (01722 429105)
	Ext. 4099
	Ext. 4122
	Dr Layth Alsaffar

Postal address:

Department of Microbiology Salisbury District Hospital NHS Foundation Trust Salisbury SP2 8BJ

DX address: Salisbury PHL DX 6930100 Salisbury 90 SP

5. Opening hours, Clinical advice, and results

5.1 Laboratory Opening Hours

The laboratory is opened:

Laboratory	
Routine: Monday to Friday	08:00-20:00
On call: Monday to Thursday	20:00-08:00
Saturdays, Sundays & Bank Holidays	09:00-17:00
Friday on call	20:00-09:00
Saturday on call	17:00-09:00
Sunday on call	17:00-08:00
РОСТ	20:00-06:00

5.2 Clinical advice

For clinical advice on diagnosis and the interpretation of Microbiology results, use of antimicrobials or infection control, Consultant advice is available:

Consultant Microbiologist	
Routine: Monday to Friday	09:00-17:30
On call Monday to Friday	17:30-09:00
Saturdays, Sundays & Bank Holidays	All day via switchboard

For advice during normal working hours: Telephone 01722 429099 (Ext: 4099) or bleep 1967.

Friday 17:00hrs to Monday 09:00hrs (non-Bank holiday weekends):

There is a cross-cover provision with Microbiology colleagues from Dorchester.

NOTE: Hospital staff – **DO NOT** use the internal bleep 1967 outside Monday to Friday (i.e., out-of-hours, weekends, and bank holidays) as this will NOT be answered.

5.3 Urgent samples

If a result is required urgently and the sample will arrive during normal working hours, the laboratory MUST be notified by telephone so that we can prioritize the request.

Please ensure that the requesting doctor's contact details are provided on the request form to enable the result to be telephoned to the requesting clinician.

5.4 Testing out of hours

The on-call service is available outside of normal Laboratory opening hours.

The Microbiology out of hours service is an urgent service. Urgent samples out-of-hours should not be sent without agreement with the laboratory on-call staff.

Once the sample has been taken, please contact the duty Biomedical Scientist: Telephone 01722 336262 (Switchboard) and give them details of the sample to be tested. Samples should be taken to the Blood Issue room (Blood bank) on level 3 and placed in the urgent sample box (Microbiology) or placed in the urgent sample box at the reception in Laboratory Medicine.

This service is restricted to only samples that require result before the next routine session. In general, samples normally accepted for the on-call service would include:

- Cerebrospinal fluid (CSF)
- Other samples approved by Consultant Microbiologist

Non-urgent samples (except blood cultures) dispatched out of hours can be placed in the microbiology refrigerator in the blood-bank room in Pathology on level 3, North Block.

Blood cultures taken out of hours should be left at room-temperature in the 'Microbiology' box in the same area.

5.5 **Requesting Tests**

All routine tests provided by the microbiology laboratory are detailed in Sections 13. All samples must be accompanied with a request form.

Amendments and addition of extra tests to initial requests can still be discussed with the laboratory after processing has started. In general, additional tests must be requested within 48 hours of sample receipt by the laboratory. In some instances, additional tests may not be possible, and a fresh specimen will be required. Further advice can be obtained from the laboratory.

Active Date: 13/06/2025

5.6 Results

Pathology results are available on the Hospital Review system or via GP computer systems immediately after authorisation.

All laboratory results are returned to the requesting clinician who has ultimate responsibility for ensuring that all results are actioned and communicated to the patient as appropriate.

In cases of difficulty or further clarification, the laboratory enquiry telephone number is 01722 429099 (Ext: 4099).

Please note that we need to establish the caller's identity before giving results over the telephone. For reasons of confidentiality (Caldicott) and Clinical Governance, we are not permitted to give results directly to patients or their relatives.

We advise all healthcare workers NOT to ask for results pertaining to themselves, but to obtain test results from the requesting physician, their doctor or from Occupational Health as appropriate.

Please NOTE: We request that users do not phone the lab to confirm whether samples have been sent or not, as this takes up much valuable time and prevents lab staff from completing their work in a timely fashion. We recommend that patient notes are annotated to confirm samples requested and taken.

5.7 **Telephoned results**

Results of urgent requests and results which may aid the immediate patient management will be telephoned. This includes all positive blood cultures, positive CSFs, and other clinically significant results.

All other results will only be telephoned on request.

5.8 **Turnaround times**

The laboratory continually monitors its turnaround times to ensure that it complies with its responsibilities within the patient pathway. The laboratory measures its turnaround times as the time from receipt until the point at which the result is authorised.

The expected turnaround times for each test are indicated on the individual test sheets in section 13 below.

5.9 Measurement of Uncertainty

Any test/procedure performed in the laboratory may be subject to a variety of factors that may influence the outcome of the test. These may occur at any of three stages:

Pre-examination

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- Examination stage
- Post-examination

However, our processes are risk assessed to reduce or mitigate adverse outcome of examinations performed.

6 Sample Collection

6.1 Viral and bacterial serology tests

A 4mL yellow top vacutainer tube is usually adequate for up to three viral serology screening tests and a referral to the reference laboratory if the screening test is positive.

For four or more tests will require two 4mL samples (2 x 4mL yellow top vacutainer). For unusual or "send away" tests not performed at SDH, an additional sample is advised to speed up handling and packaging.

Please use only the appropriate request on T-quest or Microbiology request forms for viral & bacterial serology tests. [See image 3 below].

6.2 Optimum time of and conditions for collection

Samples for bacterial culture, wherever possible, should be collected prior to commencement of antibiotic treatment.

To avoid contamination of a specimen during collection, an aseptic technique must be used: always use universal precautions, wash hands, and wear appropriate personal protective equipment. Decontamination of the sampling site or equipment may be necessary e.g., skin antisepsis before taking blood cultures or Cerebro-Spinal Fluid (CSF), or catheter port antisepsis before collecting a specimen of urine via a catheter (CSU).

Specimens must be collected into sterile containers with close fitting lids. The specimen must be clearly labelled. Once collected, place the specimen into a plastic specimen bag and seal the bag. Wash your hands and dispose of clinical waste into a clinical waste collection bag. Sharps must be disposed of safely and appropriately.

6.3 Health and safety issues pertaining to sample collection

Every specimen sent for microbiology examination should be treated as potentially infectious and standard precautions must always be observed.

When performing procedures that are likely to produce aerosols, cough inducing procedures or lancing of abscess on patients known to be infected with high-risk pathogens; appropriate PPE such as face masks, goggles or full facial visors must be worn. Used sharps must be disposed of according to Trust policy.

Samples likely to contain high risk pathogens as described by the Advisory Committee for Dangerous Pathogens are handled at a higher containment level to safeguard both laboratory staff and other downstream workers. The information is also of benefit to the patient ensuring that appropriate testing is performed.

Refer to appropriate Trust policies for further information:

• Hand Hygiene Policy

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- Infection Control policy
- Policy for the Prevention of Sharp Injuries •

7 Sample Containers

Supply of sample containers 7.1

The following Microbiology consumables can be obtained from the following Materials Management Team, contactable on: Sft.pathstores@nhs.net

Consumable	Description
	Handwritten request form (For locations that do not have access to TQUEST only)
	Container with boric acid - for urine bacteriology specimens
Urine Z	Container without boric acid - for urine bacteriology specimens
	Bacteriology swabs in Amies transport medium with charcoal
	Bacteriology swabs in Amies transport medium
35	Pernasal swab for whooping cough
	Viral swabs in viral transport medium
	Faeces container
	Sterile universal container
	Sputum container

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23 Mill monocola	Cobas transport media for <i>C</i> . trachomatis and <i>N. gonorrhoeae</i>
	Blood culture bottles
	Vacutainer tubes for blood samples
	Urgent nasopharyngeal swab for Covid-19
PERMARA BEA	Vacutainer tube for blood samples (Peach pink top, paediatric EDTA sample tube)
Serie	Sterile plastic bijoux container
	Fungal culture kit

7.2 Selection of appropriate containers

Please see Repertoire of Tests Section 13 for the selection of appropriate container for test.

Sample containers must be CE marked. Specimen containers must be leakproof and be sufficiently robust to withstand stresses during transit. Only containers approved by the Microbiology Department may be used to ensure sample integrity during transit to the Laboratory. Samples sent in non-approved containers may not be processed by the Laboratory. It is the responsibility of the person sending the sample to the Laboratory to ensure that the container used for transportation is appropriate.

The container must be adequately closed to avoid leakage. Samples that have leaked in transit may not be processed by the Laboratory.

NOTE: Users are reminded to only retain sufficient stock of sample containers for normal usage and to check the **expiry date** of stock on a regular basis and before giving out to patients or collecting sample with it.

Active Date: 13/06/2025 Review Date: 13/06/2027 CONTROLLED DOCUMENT; UNCONTROLLED WHEN PRINTED OR COPIED Page 13 of 82

Document Title: Microbiology User Handbook Q-Pulse Index: MIC-POL-15 Revision Number: 2.1 Author: Olutoye, Yinka

7.3 Labelling of sample containers

Clinical governance requires that sample container is labelled with sufficient information to provide an unequivocal link with the request form and the patient from whom they are collected.

Pre-printed addressograph labels are acceptable on sample containers. However, they must contain all necessary patient details.

Minimum data set for identification:

- Patient's surname
- Patient's forename
- Date of birth and/or hospital number/NHS number

Microbiology sample containers should additionally include type of sample and/or site of collection.

NOTE: Multiple samples taken at different times, sample container MUST be labelled with the time the sample is taken (24 hr clock).

NOTE: Failure to label sample containers appropriately may result in the sample being rejected.

8 Request Forms

All samples must be accompanied by a properly completed request form. Request forms MUST contain sufficient information to provide an unequivocal link with the sample and the patient from whom they are collected.

The following patient detail MUST be added written on the request form:

- Patient's surname
- Patient's forename
- Date of birth and/or hospital number/NHS number

In addition to the above details, the following should be noted:

- Tests for antibiotic assay levels, must contain the following details:
 - Mg of last dose given
 - Date and time of last dose
 - > Date and time when sample was taken.
- Serology requests should include onset date of symptoms as this has relevance to interpretation of results.

All requests forms MUST include the requesting physician's signature.

DO NOT add a microbiology tests, (e.g., viral serology), to a Laboratory Medicine sample/form. This may cause serious delays in the sample arriving at the laboratory and result in insufficient sample for testing.

NOTE: Failure to comply with the above guidance may result in the sample being rejected

8.1 Electronic requesting

Please use electronic requesting (T-QUEST) orders where available. It is important to ensure that the correct sample accompanies the correct request form before placing inside the sample bag.

Please ensure that you order the correct test and select the correct sample type as failure to do so may lead to incorrect/none testing. If the test you require is not visible, please contact the laboratory to check that the test is available.

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Include clinical details and symptoms, as well as information on antibiotic use, foreign travel, outbreaks, date of onset, etc.

Where TQUEST requesting is not available handwritten request forms must be used.

			Sumame	Forena		
	85032		EDITESTPATIENT NHS Number	Date C	TYNINE	Sex
		· · · · ·	Fasting	13/01/ Pregna		Fema
			Location Acute Medical Unit	Consu Cullis		Categ
	#TQI1429941	1	Patient Address	[Cuilis	30	Copy
Date/Time Of Coll	lection		Tel:			
			Requester Name		al Number	Urgen
Specimen Type	101.0	-	Anthony Wood Antibiotics	11112	5	No
Blood culture bo Site Of Specimen		*				
Blood from perip						
Investigation Desc			Collected by (signature):			
Blood cultures Clinical Details			-			
Test test test test	it test					
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Image 2: Microbiology TQuest form

Document Title: Microbiology User Handbook Q-Pulse Index: MIC-POL-15 Revision Number: 2.1 Author: O



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[Lab Use Only] Affix lab sample ID label here		Sumame TEST	Forename ANDY		
		NHS Number	Date Of Birth 12/04/1957	Sex Unspecified	
		Fasting	Pregnant		
		Location Farley	Consultant Cullis J O	Category NHS	
#TQI3676480 Date/Time Of Collection		Patient Address SAL DIST HOSP Tel:		Сору То	
Specimen Type Blood Serum (SE) (Gold Top SST)		Requester Name James Ryan	Hospital Number TEST	Urgency No	
Sideo Carlan (C2) (2004 Top C01) Investigation Description Zika virus serology Clinical Details TEST TEST		Date of onset of symptoms			
		Collected by (signature)			
		1	e	1. A.	

Serology -- IR Requesting -- Salisbury NHS Foundation Trust

*** REMEMBER ***

HIGH RISK specimens from patients with known or suspected infection with TB or serious exotic infections are hazardous. Use 'Danger of Infection' labels on the specimen container, request form and plastic bag.

	Surname TEST	Forename ANDY	Г	
[Lab Use Only] Affix lab sample ID label here	NHS Number	Date Of Birt 12/04/1957		MEY TEST
	Fasting	Pregnant		12/04/1957 TEST
	Location Farley	Consultant Cullis J O	Category NHS	#TQ:0676480
#TQI3676480 Date/Time Of Collection	Patient Address SAL DIST HOSP Tel:		Сору То	AMDY TEST 12/04/1957 TEST
Specimen Type Blood Serum (SE) (Gold Top SST)	Hospital Number TEST	Urgency No	-	#TCJ3676480
Site Of Specimen / Qualifier	Date of onset of sympton	ns		MEY TEST
nvestigation Description Zika virus serology	Collected by (signature):	Collected by (signature)		
Clinical Details				
TEST TEST TEST				MDN TEST 12/04/1957 TEST #TQ/36/76460
				ANDY TEST 1204/1957 TEST #TCI00/06400

Image 3: Serology TQuest Form

8.2 Manual requesting

In the absence of an electronic T Quest form, a handwritten form can be used. When completing a handwritten form, ensure to fill the following details correctly.

Minimum Data Set for Identification:

- Hospital number and/or NHS number
- Patient surname and forename (in full, not initials)
- Date of birth (DOB)
- Patient address if hospital number / NHS number not supplied.

In addition to the minimum data set for patient identification, ensure all other relevant fields are completed such as:

- Ward/ Practice, Consultant/GP
- Patient address
- Patient gender
- Date and time of collection
- Specimen type Active Date: 13/06/2025

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Author: Olutoye, Yinka

- Investigation(s) required
- Name of requesting clinician and bleep number
- Relevant clinical details
- Current drug therapy
- Copy reports, if required
- Patient category (PP/NHS)

To ensure samples can be safely and appropriately tested in the laboratory, include details of foreign travel, symptoms and known or suspected contact with other patients known to have communicable disease.

It is essential to use a ballpoint pen when completing request forms and ensure that the same details are on both top and bottom copies of the request form.

DO NOT use a felt tip and fountain pen to fill the form. This can lead to delay in processing samples, or requests being missed altogether, as carbon copies are often incomplete. When addressograph labels are used, please ensure that a label is fixed to EACH part of the request form.

The second se				SPECIMEN:	Date	Tim
NHS No.	111					
Surname	TII	1111		Investigation Required:		
Forename					EASE WRITE LEGIBL	
Sex F	Date of	Birth		Clinical Details: Samples lacking relevan	t information canno	ot be proces
Hospital Reg	No.	Consultant / G	.P. Location / W	ard		
	Category 2	Private Cop	y to:			
Patient's add	ress					
ANTIBIOTIC	THERAPY (indi	cate recent, curren	t or intended)			
SAL009						
13				Date of onset:		

Image 4: Handwritten Request Form

8.3 Anonymous/uniquely identified samples

In certain circumstances patient identification details are intentionally hidden or substituted with particular identification numbers (e.g., Sexual Health patients). In such instances, a properly coded identifier must be used in place of the patients last name and first name (e.g., GUM patient samples need to have a GUM number, patient gender, and DOB).

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9 Sample Acceptance and Rejection Criteria

Sample acceptance criteria ensure adequate identification for Microbiology samples and request forms for them to be accepted by the laboratory for analysis. It is important to clearly identify the investigations required with relevant supporting information. Inadequate or inaccurate labelling might cause delays before Microbiology results are available and affect patient care.

In case the sample is not repeatable or not reproducible, it will be processed but will have a disclaimer added onto the report. Microbiology will accept no responsibility for samples analysed which initially failed to meet the acceptance criteria. Where the sample is repeatable/ reproducible, no analysis will be performed, and an appropriate comment will be included on the Microbiology report.

Samples and request form must be received with all required matching patient details. This must be received in a safe condition i.e., not leaking/stained with bodily fluids as this can cause a health risk to transport staff, and laboratory staff alike.

The Microbiology Laboratory will reject any sample in the following scenarios:

- Sample with no unique identification of the patient i.e., they do not meet the minimum data set for identification,
- An incorrect sample type has been received for the test(s) requested,
- Sample is received in a condition that poses health and safety risk to staff e.g., leaking or sharps attached,
- Sample or request form is unlabelled or incorrectly labelled with less than the minimum data sets for patient identification,
- There is a mismatch of details between the form and sample(s) received,
- The information provided is illegible,
- Collection container is out of date.

Where possible the requester will be contacted by telephone and advised of the reason for the sample being rejected (and a repeat where possible being sent). A rejected sample will result in a report indicating the key reasons for rejection, with a request for a repeat sample being included where appropriate.

10 Transportation of Samples

Please refer to the Trust Specimen Transportation Policy for the correct procedures for submitting samples to the laboratory.

10.1 Transportation of routine samples to the laboratory

All specimens must be transported in a timely manner such that it preserves the integrity of the sample and allows for rapid testing in urgent situations.

10.1.1 Pneumatic Chute

Samples may be sent direct to the laboratory using the Pneumatic chute (Whooshy) system:

- Pathology address: 900
- Microbiology address: 901

NOTE:

- DO NOT use the vacuum transport tube (whooshy) to transport high-risk samples.
- DO NOT use this system when the laboratory is closed i.e., out-of-hours.

Active Date: 13/06/2025

Review Date: 13/06/2027

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• DO NOT use to transport CSF sample.

10.1.2 Portering Services

During routine working hours the theatre porters routinely deliver samples to pathology department three times daily direct to level 4: 08.30, 12.30, and 16.40.

Specimens from theatres, DSU and New Hall are delivered to level 4 and signed for by laboratory staff.

10.1.3 Courier Services

The hospital couriers collect samples from external clinics, other outlying hospitals, and GP surgeries. These samples are transported in **UN3373** approved carriers with secure and leak proof lids that can be closed and sealed. The boxes carry a warning label and are lined with absorbent material to absorb any leakage. Samples are delivered to Laboratory Medicine on Level 3.

There are ten transport deliveries each day and these arrive between approximately 11.00am and 19.00pm.

Where this is not practicable due to delays in transportation samples should be kept refrigerated. Samples may be kept in a refrigerator at a temperature of 4-8°C for a maximum of 24 hours prior to transportation.

NOTE: Blood Culture bottles MUST NOT be refrigerated. These must be transported to the Laboratory as soon as possible for incubation at 37°C within 3 hours of collection.

Out-of-hours (from 20:00 until 08:00 Monday to Friday, from 17:00 until 09:00 on Saturdays and Bank Holidays and all-day Sunday) the vacuum tube to the Microbiology reception is switched off, and any samples sent may be randomly sent to locations other than the laboratory!

NOTE: For transportation of samples to the laboratory from external sites or by post, and use of the pneumatic chute system, please refer to the Trust Specimen Transportation Policy. In cases of difficulty or further clarification, the laboratory enquiry telephone number is 01722 429099 (Ext: 4099).

10.2 Transportation of urgent samples

Urgent samples must be sent to the laboratory immediately. To discuss an urgent sample with the oncall Biomedical Scientist out of hours please phone Switchboard: Telephone 01722 336262 and ask for the on-call Biomedical Scientist.

10.3 High Risk Samples

All samples should be regarded as potentially infectious. However, certain samples constitute a potential higher risk of infection to persons handling them:

- Typhoid/paratyphoid fever
- Dysentery
- Tuberculosis (samples from sites where tuberculosis infection is likely)
- Anthrax
- Brucellosis
- Transmissible Spongiform Encephalopathy (including CJD)
- Viral haemorrhagic fever

Active Date: 13/06/2025



- Avian Flu
- MERS

To transport such samples to the laboratory, please refer to the Trust's Policy for the Transport of Pathology Specimens.

To minimise the risk of infection from these samples, ensure they are packaged as follows:

- Flag as "Danger of Infection" on Tquest,
- Specify the nature of the risk on the request form,
- Use unambiguous and commonly recognised terminology,
- Place the sample in a sealable plastic bag and close the seal.

This is a necessary procedure to safeguard both laboratory staff and other downstream workers. **Note:** The Consultant Microbiologist <u>**MUST**</u> be contacted <u>**BEFORE**</u> collecting specimens from a patient suspected of having a Viral Haemorrhagic Fever, Human Avian Flu or CJD.

11. Sample storage

See the table below for sample retention times. If additional investigations are required, please contact the laboratory.

Type of Clinical Material	Retention time
Semen-vasectomy	Kept for 48hrs after result is completed
Semen- infertility	
Urine samples	3 days
Covid-19 samples (NEGATIVE)	4 days
Bacteriological swabs Sputum/NPA/BAL samples	1 week
High risk/Danger of infection samples	
Faeces samples	
Rejected samples	
Chlamydia samples	
Mycology samples	3 weeks
Tissue/Fluid samples	1 month
CSF	Kept for current month plus 1 month
Resistant organisms from urine samples	3 months
MRSA and resistant organisms from other samples	
Covid-19 samples (POSITIVE)	Up to 6 months (as storage allows)
C. difficile positive samples	1 year
Significant blood culture isolates	
Antenatal booking blood serum samples	2 years
Needlestick injury serum samples	
Serum samples	
Referral CL2 slopes	Kept until Reference laboratory report is
Referral CL3 Slopes	returned

12. Quality Assurance in Microbiology

The Microbiology department is committed to the provision of high-quality and professional service to our service uses. We maintain assurance of the quality of our service through the following.
Active Date: 13/06/2025
Review Date: 13/06/2027

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Document Title: Microbiology User Handbook Q-Pulse Index: MIC-POL-15 Revision Number: 2.1 Author: Olutoye, Yinka 12.1 UKAS accreditation



The laboratory is working towards accreditation by the United Kingdom Accreditation Services (UKAS).

12.2 External Quality Assurance (EQA)

The laboratory takes part in different EQA schemes from various providers to cover most of the tests provided; we participate in EQA from UK NEQAS (National External Quality Assessment Scheme), QCMD (Quality Control for Molecular Diagnostics), BMSMicro and LabQuality.

The EQA schemes enables us to compare our results with that of other laboratories using the same, or different platforms and methods.

12.3 IQC (Internal Quality Control)

We use independently sourced control materials which gives results in the expected range to control our reagents and platforms. Patient results are not released when there is IQC failure.

12.4 IQA (Internal Quality Assurance)

This is a programme of duplicate testing of samples received. The success of our IQA proves the repeatability of the results. Any significant deviation is investigated.

12.5 Acceptance testing

This is a process that ensures that new lots of reagents and kits are performing as expected before being introduced for us. We also perform suitability checks on all suppliers (including the referral laboratories), to ensure they offer a high quality, certified or accredited service.

12.6 Audits

All aspects of the laboratory work are audited bi-annually with the Quality Management System and areas of high-risk audited annually. Non-conformances are investigated and suitable corrective actions are put in place to prevent future occurrence.

12.7 Quality indicators

Microbiology department also have quality indicators that are used to measure the quality of work done, the include but not limited to:

- **Turnaround Times** these are monitored monthly to ensure patient care is adequately and efficiently met.
- **Training and Competence** all our staff are adequately trained, and competency assessed to carry out their task. Our staff are encouraged to participate in Continuous Professional Development which is supported by event provided by the Trust.
- User Feedback- Feedback from our users is sort annually and reviewed to enable us to address issues and concerns from them. Complaints and concerns are investigated and responded to within timescales found in Trust guidelines.

Active Date: 13/06/2025



Author: Olutoye, Yinka

- **Datix Incident Reporting**-Incidents are recorded through the Datix Incident Reporting form of the Trust. All incidents are investigated, and appropriate actions put in place.
- **Staff Suggestions** Staff are encouraged to contribute to quality in the department. All range of staff grades have mediums of adding value to the service we provide.

13. Repertoire of Tests (A-Z)

This section covers the tests that our laboratory offers according to the service repertoire agreed with our users.

Our laboratory offers a range of specialist tests which are undertaken at reference laboratories. The information where a specific specialist test is performed is indicated in the 'Additional Comments' section for each test.

Please contact the laboratory for any queries regarding the tests offered and information regarding any special sample requirements.

Please find a test using the index.

13.1 Bacterial Culture, Fungal Culture and Viral Investigations

Blood culture

Investigation	Blood Cultures			
Tests	Gram stain, if positive, Culture & Sensitivities			
Sample type	Adult: 5-10 ml of blood per bottle.			
	Paediatric: 3-4 ml of blood per bottle.			
Collection Container	Adult Blood Culture set: Aerobic (blue) and Anaerobic (purple) bottles.			
	Paediatric Blood Culture bottle: Aerobic (yellow) bottle.			
Sample collection	 Samples should be collected before antimicrobial treatment. 			
	Samples should be taken as soon as possible after a spike of			
	fever.			
	N.B. To guarantee optimal result please ensure that the blood culture set			
	is transported to the laboratory within 3 hours of the sample being			
	collected.			
Turnaround Time	36 hours- interim paediatric result			
	5 days- Negative result			
	Positive results are phoned to the wards as soon as possible.			
	>5 days if the sample requires extended culture.			
Limitations	Blood cultures should only be taken when there is a reason to			
	suspect infection and not for routine assessment.			
	 Delays in transportation may affect the recovery of pathogens. 			
Active Date: 12/06/2025	Paviow Date: 12/06/2027			

Active Date: 13/06/2025



Revision Number: 2.1	Author: Olutoye, Yinka	
	 Any recent antimicrobial therapy can have a significant effect on blood culture results by decreasing the sensitivity of the test. If patients have received previous antimicrobial treatment, bacteraemia should be considered even if blood culture results are negative. False negatives may occur if inadequate blood culture volumes are submitted. 	
Out-of-Hours-Testing	Bottles should be left at room temperature in blood-issue room.	
	DO NOT PLACE IN THE FRIDGE	
Additional Comments	DO NOT REMOVE OR COVER UP THE BARCODE LABELS AS THESE ARE	
	REQUIRED IN THE LABORATORY.	

Bordetella pertussis

*	
Investigation	Bordetella pertussis culture
Tests	Isolation and characterisation of Bordetella species.
Sample type	Pernasal swab
Collection Container	Pernasal swab (blue top)
Sample collection	 A pernasal swab is inserted through a nostril and advanced along the floor of the nose until it reaches the nasopharynx. Optimally collected before antimicrobial therapy started. Specimens should be sent to the laboratory without delay during normal working hours. Outside of normal working hours samples should be refrigerated. NOTE: Delays of over 48 hours are undesirable.
Turnaround Time	7 days
Limitations	 Samples taken >2 weeks after onset of symptoms may not yield a positive result. It is advisable to also rule out respiratory viruses when testing for pertussis. Two sample should be taken: one for the culture of Bordetella species and the other for respiratory virus PCR. If pertussis PCR is required, send a dry swab i.e., swab without transport medium. Delays in transportation may affect the recovery of pathogens.
Out-of-Hours-Testing	No
Additional Comments	None

Candida Antifungal Susceptibility testing

Investigation	Antifungal susceptibility testing for Candida species.
Tests	Antifungal susceptibility testing
Sample type	Pure culture
Collection Container	N/A
Sample collection	N/A
Turnaround Time	10-14 days

Active Date: 13/06/2025

Document Title: Microbiology User Handbook Q-Pulse Index: MIC-POL-15



Revision Number: 2.1	Author: Olutoye, Yinka
Limitations	Consultant Microbiologist will specify which drugs are to be tested.
Out-of-Hours-Testing	No
Additional Comments	This test is performed at UKHSA Bristol

Cerebro-Spinal Fluid (CSF) Culture

Cerebro-spinal Fluid (CSF) Culture	
Cell count, Gram stain and Culture	
1–2ml CSF	
Two sterile plastic bijoux containers.	
Please send 1st and 3rd samples.	
 Optimally collected before antimicrobial therapy started. Do not delay antibiotic administration if clinically indicated. 	
 Dispense CSF (minimum 0.5ml in each bottle) into single use 	
containers and label in order.	
 Specimens should be sent to the laboratory without delay during normal hours. 	
 Outside of normal hours samples should be placed in the 	
pathology reception and the on-call Microbiology Biomedical Scientist contacted through switchboard.	
 State if TB culture or Cryptococcal culture/antigen testing is required. 	
2 hours for microscopy.	
3 days for culture.	
 Cells degenerate. A delay in transportation may produce a false cell unrepresentative of the clinical picture and may affect the 	
recovery of pathogens.	
Yes, by arrangement, 24/7.	
None	

Corneal Scrape Culture

Investigation	Corneal Scrape Culture
Tests	Gram stain & Culture
Sample type	Corneal scrape
Collection Container	Direct inoculation onto plates (CBA, CHOC, FAA & SAB), slide and into saline bottle (if Acanthamoeba investigation is required)
Sample collection	Optimally collected before antimicrobial therapy started.
	Carefully smear material onto agar plates and onto glass slide
	(circle area with permanent marker).
	 If insufficient specimen to make an impression smear and
	inoculate plates, please prioritise culture plates.

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Revision Number: 2.1	Author: Olutoye, Yinka	
	 Specimens should be sent to the laboratory without delay during normal working hours. Outside of normal working hours samples should be refrigerated. Delays of over 48 hours are undesirable. 	
Turnaround Time	2 hours for microscopy (During normal working hours)	
	2 – 5 days for culture.	
Limitations	 Requires good amount of cellular material. 	
	• Delays in transportation may affect the recovery of pathogens.	
Out-of-Hours-Testing	No	
Additional Comments	None	

Faecal Samples for bacterial, viral and parasitology tests

Investigation	Tests	
Faeces Culture	Culture and sensitivities	
Cryptosporidium	ZN stain for the detection of Cryptosporidium sp.	
microscopy		
Helicobacter pylori	Helicobacter pylori antigen testing	
antigen detection		
Clostridium difficile	Toxin Detection	
Toxin		
C. difficile PCR	PCR	
ribotyping		
Detection of viruses	Faecal Viral PCR	
e.g., enterovirus		
Parasitology	Microscopy	
Rotavirus EIA	Antigen detection	
Norovirus	PCR	
Collection Container	Universal with spoon (blue top)	
Sample collection	Optimally collected before antimicrobial therapy started.	
	 Specimen may be passed into a clean, dry, disposable bedpan or 	
	similar container and transferred to an appropriate collection	
	container.	
	• Specimens should be sent to the laboratory during normal	
	working hours.	
	Outside of normal working hours samples should be	
	refrigerated.	
Turnaround Time	Faecal Culture: 4 days	
	Cryptosporidium microscopy: 2 days	
	Helicobacter pylori: 1 day	
	• C. diff toxin: 1day	
	• C. diff ribotyping: 10-14 days	
	• Enterovirus: 10-14 days	
	Parasitology: 6 days	
	• Rotavirus: 1 day	
Limitations	Clinical details are essential for processing.	
	 Delays in transportation may affect the recovery of pathogens. 	
Active Date: 13/06/2025		

Active Date: 13/06/2025



Revision Number: 2.1	Author: Olutoye, Yinka
	 <i>H. pylori</i>- This is a qualitative assay for <i>H. pylori</i> antigen in stool. A negative result does preclude the possibility of infection with <i>H. pylori</i>.
	 Rotavirus- Limited to children <5 years old.
Out-of-Hours-Testing	No
Additional Comments	 Cryptosporidium microscopy is only tested on BS5-BS7 type stool samples or if specifically indicated in clinical details.
	 H. pylori antigen testing is specific and allows post treatment testing or re-testing if symptoms re-occur despite therapy.
	 <i>H. pylori</i> stool antigen testing is more specific and allows post treatment testing or re-testing if symptoms re-occur despite therapy.
	 Public Health England produces a useful guide on who and when to test for <i>H. pylori</i>:
	https://www.gov.uk/government/publications/helicobacter-
	pylori-diagnosis-and-treatment
	 C. diff toxin test is only performed on liquid / semi-formed stools BS5-BS7 type stool.
	• <i>C. difficile</i> toxin test is performed on in-patient samples, patients over 65 years old or patients with history of antibiotic-associated diarrhoea (please state 3 months antibiotic history).
	 Samples from children < 2 years old are unsuitable for investigation for <i>C. difficile</i>.
	 Do not request for <i>C. difficile</i> if a positive result is within previous 28 days.
	 Parasitology- Please contact Laboratory if 'hot-stool' examination is required.
	 Detection of viruses- is performed at UKHSA Bristol
	• C. diff ribotyping is performed at CDRN Leeds
	Cryptosporidium typing is performed at PHW, Swansea

Fungal Culture

Investigation	Fungal Culture
Tests	Microscopy & Culture
Sample type	• Skin
	Nails
	• Hair
Collection Container	Fungal culture kit
Sample collection	Skin Specimens: Skin lesions should be collected by scraping skin from
	the advancing edge of the lesion with a blunt scalpel blade or other

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Review Date: 13/06/2027

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	sharp instrument. Place the scraping into a special Mycology transport	
	pack. Please make sure you send enough material for both microscopy	
	and culture. At least 5mm of skin flakes are required.	
	Nails: Clippings should include the full thickness of the nail and extend as	
	far back from the edge as possible. Samples should be sent in a	
	Mycology transport pack. Several small parings are preferred to one	
	large sample to optimise culture results. Nail parings should be taken	
	from the diseased area, the discoloured or brittle parts of the nail and	
	cut back as far as possible from the free edge as some fungi are	
	restricted to the lower parts. Scrapings can also be taken from under the	
	nail to supplement the clipping. Nail clippings often fail to grow fungi	
	even if present.	
	Hair: Hair should be plucked from affected areas together with skin	
	scrapings from associated scalp lesions. Broken lustreless hair should be	
	selected from the margin of the scalp lesion. Hair should be removed	
	with epilating forceps. The hair follicle and one inch of proximal hair	
	should be sent to the laboratory. Receipt of cut distal ends will not be	
	processed.	
Turnaround Time	7 – 10 days for microscopy	
	3 – 4 weeks for culture	
Limitations	 Swabs are of little value for the investigation of dermatophyte 	
	infections.	
	 Delays in transportation may affect the recovery of pathogens. 	
Out-of-Hours-Testing	No	
Additional Comments	For further information and guidance on recommended treatment	
	options please refer to NICE website: Fungal skin infection - body and	
	groin Health topics A to Z CKS NICE	

Gonococcal Culture (GUM clinic only)

· · · · · · · · · · · · · · · · · · ·		
Investigation	Culture for Neisseria gonorrhoeae (GUM clinic only)	
Tests	Culture and sensitivities	
Sample type	Endo-cervical swab	
	Urethral swab	
	Throat swab	
	Rectal swab	
Collection Container	Transport swab (black top)	
Sample collection	No special requirements.	
	 Specimens should be sent to the laboratory during normal 	
	working hours.	
	NOTE: SWABS FOR GONOCOCCAL INVESTIGATION SHOULD NOT BE	
	REFRIGERATED as this significantly reduces the recovery rate.	
Turnaround Time	4 days	
Limitations	 Clinical details are essential for processing. 	
	• Delays in transportation may affect the recovery of pathogens.	
Out-of-Hours-Testing	No	
Additional Comments	None	

Active Date: 13/06/2025



Author: Olutoye, Yinka

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Investigation	Gynaecological Culture	
Tests	Microscopy (where applicable) & Culture	
Sample type	Vaginal swab	
	Endocervical swab	
	Urethral swab	
	Penile swab.	
Collection Container	Transport swab (black top)	
Sample collection	No special requirements.	
	 Specimens should be sent to the laboratory during normal 	
	working hours.	
	 Outside of normal working hours samples should be 	
	refrigerated.	
	 Optimally collected before antimicrobial therapy started. 	
Turnaround Time	4 days	
Limitations	 Essentially, in uncomplicated cases of vaginal discharge a 	
	diagnosis can be reached using clinical history, characteristic	
	appearance, and the pH of the discharge.	
	Speak to Consultant Microbiologist if culture for <i>Neisseria</i>	
	gonorrhoeae is required. The laboratory now provides PCR for	
	the detection of <i>N. gonorrhoeae</i> .	
	• For <i>N. gonorrhoeae</i> testing please send a Cobas PCR <i>Chlamydia</i>	
	swab and make it clear that testing for <i>N. gonorrhoeae</i> is	
	required. One Cobas swab can be used to test for both	
	Chlamydia and N. gonorrhoeae if requested.	
Out of Hours Testing	Delays in transportation may affect the recovery of pathogens.	
Out-of-Hours-Testing	No	
Additional Comments	None	

Infection Control Screening Tests

Investigation	Tests
MRSA	Culture & Sensitivities
MRSA	PCR
VRE	Culture & Sensitivities
CRE	Culture & Sensitivities
Candida auris	Culture & Sensitivities
Sample type	
MRSA	Swab, Urine, Sputum
MRSA PCR	Nasal swab
VRE	Rectal swab, Faeces, other swabs
CRE	Rectal swab
Candida auris	Swab, Urine, Sputum
Collection Container	Transport swab (Black topped)
	Universal (White top)

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Review Date: 13/06/2027

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	Universal (60ml wide-mouth, metal top)	
	Universal with spoon (blue top)	
	Viral swab for PCR	
Sample collection	No special requirements.	
	 Specimens should be sent to the laboratory during normal 	
	working hours.	
	Outside of normal working hours samples should be	
· · · · · · · · · · · · · · · · · · ·	refrigerated.	
Turnaround Time	Negative 1 2 days	
MRSA culture	Negative: 1 – 2 days Positive: 2-4 days	
MRSA PCR	1 day	
VRE	Negative: 1 – 2 days	
	Positive: 2-4 days	
CRE	Negative: 2 days	
	Positive: 4 days	
Candida auris	Negative: 1 – 2 days	
	Positive: 2-4 days	
Limitations	Delays in transportation may affect the recovery of pathogens.	
Out-of-Hours-Testing	No	
Additional Comments	See Trust's Infection Control Policies	

Intravenous (IV) Cannula Culture, e.g., CVP line tip

Investigation	IV Cannula Culture, e.g., CVP line tip
Tests	Culture
Sample type	End of cannula tip (end 4 cm)
	Note: blood culture is preferable
Collection Container	Universal (white top)
Sample collection	Sample should be collected before antimicrobial therapy started.

Active Date: 13/06/2025



infection is NOT suspected.

Neonatal Screen Culture

Investigation	Neonatal Screen Culture
Tests	Culture
Sample type	Swabs
	Gastric aspirate
Collection Container	Transport swab (black top)
	Universal container
Sample collection	No special requirements.
	 Specimens should be sent to the laboratory during normal working hours.
	 Outside of normal working hours samples should be refrigerated.
Turnaround Time	4 days
Limitations	Delays in transportation may affect the recovery of pathogens.
Out-of-Hours-Testing	No
Additional Comments	

Parasitology investigation for Enterobius vermicularis

Investigation	Microscopy for Enterobius vermicularis
Tests	Microscopy
Sample type	Sellotape slide
	Perianal swab.
Collection Container	Laboratory no longer supplies collection kits.
	Sellotape slide
	Cotton-wool swab in dry container.

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Revision Number: 2.1	Author: Olutoye, Yinka
Sample collection	Sellotape slide
	Apply clear Sellotape to the perianal region, pressing the adhesive side
	of the tape firmly against the left and right perianal folds several times;
	the tape can be wrapped around a tongue depressor to aid specimen
	collection. Smooth the tape back on the slide, adhesive side down.
	Perianal swab
	Perianal specimens are best obtained in the morning before bathing or
	defecation. Three specimens should be taken on consecutive days
	before pinworm infection is ruled out. Cotton-wool swab in dry
	container should be used for collection.
	Spread buttocks apart and rub the moistened cotton wool swab over the
	area around the anus, but do not insert into the anus. Place cotton wool
	swab back in its container (no transport medium required).
	Occasionally, an adult worm may be collected from a patient and sent in
	saline or water for identification.
	Specimens should be sent to the laboratory without delay during normal
	working hours.
	Outside of normal working hours samples should be refrigerated.
Turnaround Time	6 days
Limitations	
Out-of-Hours-Testing	No
Additional Comments	

Pus Culture

Investigation	Pus Culture
Tests	Gram stain, Culture & Sensitivities
Sample type	Pus
Collection Container	Universal (white top)
Sample collection	 No special requirements. Specimens should be sent to the laboratory during normal working hours. Outside of normal working hours samples should be refrigerated.
Turnaround Time	5 days
Limitations	Delays in transportation may affect the recovery of pathogens.
Out-of-Hours-Testing	No
Additional Comments	None

Respiratory culture

Investigation	Sample type
Broncho-alveolar	Broncho-alveolar lavage
lavage Culture	
Pleural Fluid Culture	Pleural Fluid
Sputum Culture	Sputum
Tests	Gram Stain (where applicable), Culture
Collection Container	Universal (white top)

Active Date: 13/06/2025

Review Date: 13/06/2027

Salisbury



	Sputum container
Sample collection	 Specimens should be fresh and optimally collected before antimicrobial therapy started. Minimum sample size is preferably 5mL. Specimens should be sent to the laboratory without delay during normal working hours. Outside of normal working hours samples should be refrigerated. Sputum: If fungal culture required e.g., in an immuno-compromised patient, please indicate on request form
Turnaround Time	Sputum: 4days BAL: 5 days Pleural Fluid: 5 days
Limitations	 Delays in transportation may affect the recovery of pathogens. Any delay may allow overgrowth of Gram-negative bacilli, and <i>Haemophilus</i> species and <i>Streptococcus pneumoniae</i> may not get isolated.
Out-of-Hours-Testing	No
Additional Comments	 Contact Consultant Microbiologist if <i>Pneumocystis jirovecii</i> testing is required. BAL samples are suitable for overnight refrigeration only, they must not be stored over a weekend.

Staphylococcus and Streptococcus Reference Service (and PVL)

Investigation	Staphylococcus and Streptococcus Reference Service including PVL
	testing
Tests	Genotyping, Toxin detection, etc.
Sample type	Pure culture
Collection Container	N/A
Sample collection	N/A
Turnaround Time	10-14 days
Limitations	Consultant Microbiologist will decide which isolates need to be sent for
	further investigation.
Out-of-Hours-Testing	No
Additional Comments	These tests are performed at UKHSA Colindale (BRD)
	NOTE: Sample will need to be sent for culture and sensitivity before
	isolate can be sent for PVL testing

Sterile Fluid Culture

Investigation	Sample type

Active Date: 13/06/2025

Document Title: Microbiology User Handbook Q-Pulse Index: MIC-POL-15 Revision Number: 2.1 Author: (



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Revision Number: 2.1	Author: Olutoye, Yinka
Ascitic Fluid Culture	Ascitic Fluid
Joint Fluid Culture	Joint Fluid
Tests	Gram stain and culture, Crystal (Joint fluid)
Collection Container	Universal (white top) (For direct culture and microscopy)
Sample collection	 Optimally collected before antimicrobial therapy started. Samples should be taken using strict aseptic technique by trained medical staff in line with Trust procedure. Ideally a minimum volume of 1ml should be collected. Where adequate, sample can also be inoculated into blood culture bottle set. Specimens should be sent to the laboratory without delay during normal working hours. Outside of normal working hours samples should be refrigerated. Joint Fluid-Ideally a minimum volume of 1ml should be collected. Ascitic fluid-Where there is adequate sample, inoculate into blood culture bottle set.
Turnaround Time	 Ascitic fluid-4 days, 5 days if sent in Blood Culture bottles Joint fluid-5 days
Limitations	Delays in transportation may affect the recovery of pathogens.
Out-of-Hours-Testing	No
Additional Comments	Please indicate if microscopy for crystals is required.

Swabs for Microscopy, Culture and Sensitivities (M, C &S)

Investigation	Sample Type
Ear Swab Culture	Ear swab
Eye swab culture	Eye swab
Mouth swab culture	Mouth swab
Nose swab culture	Nose swab
Throat swab culture	Throat swab
Skin swab culture	Skin swab
Wound swab culture	Wound swab
Ulcer swab culture	Ulcer swab
Tests	Microscopy, Culture & Sensitivity
Collection Container	Transport swab (black top)
Sample collection	No special requirements.

Active Date: 13/06/2025

Document 1 Q-Pulse Inde **Revision Nu**

Document Title: Microbiology User Handbook	
Q-Pulse Index: MIC-POL-15	Salisbury NHS Foundation Trust
Revision Number: 2.1	Author: Olutoye, Yinka
	 Specimens should be sent to the laboratory during normal working hours. Outside of normal working hours, samples should be refrigerated.
Turnaround Time	4 days NOTE: Throat swab can take longer if prolonged culture is required in the case of persistent/recurrent sore throats
Limitations	 Delays in transportation may affect the recovery of pathogens.
Out-of-Hours-Testing	No
Additional Comments	 DO NOT USE FOR <i>N. GONORRHOEAE</i> AND/OR <i>C. TRACHOMATIS PCR</i> DO NOT USE FOR HERPES SIMPLEX PCR Nasal swabs should NOT be taken for the investigation of <i>Bordetella pertussis</i>, see <i>Bordetella pertussis</i> above. Isolation of <i>Neisseria</i> spp. from Throat swab is only on request. Pus sample should be sent in a white topped universal instead of pus swab. For leg ulcers, only send swabs if there is clear evidence of infection, e.g., spreading erythema around the ulcer, new

	pus, cellulitis, increasing pain.	
٠	Before sampling remove colonising organisms by washing	
	with sterile saline. Use swab to get deep to the ulcer base	
	and under any over-hanging edges.	

Provide description of any clinical signs to aid interpretation • of results.

Tissue Culture

Investigation	Tissue Culture
Tests	Gram stain, Culture & Sensitivities
Sample type	Tissue
Collection Container	Universal (white top)
Sample collection	No special requirements.
	 Specimens should be sent to the laboratory without delay during normal working hours.
	 Outside of normal working hours samples should be refrigerated.
Turnaround Time	7 days
Limitations	Please do not send samples in formalin.
	• Delays in transportation may affect the recovery of pathogens.
Out-of-Hours-Testing	No
Additional Comments	



Author: Olutoye, Yinka

TB Culture (Urine)

Investigation	TB Culture (Urine)
Tests	Culture
Sample type	First-pass early morning urine (from 3 consecutive days)
Collection Container	Universal (60ml wide-mouth, metal top)
Sample collection	• 3 specimens should be sent from 3 consecutive days.
	 Specimens should be sent to the laboratory during normal working hours.
	 Outside of normal working hours specimens should be refrigerated.
Turnaround Time	6 weeks
Limitations	N/A
Out-of-Hours-Testing	No
Additional Comments	NOTE: No microscopy performed on urine TB samples This test is
	performed at University Hospital Southampton.

TB Culture (Sputum/ BAL/ Tissue/ Pus)

Investigation	TB Microscopy and Culture (Sputum/ BAL/ Tissue/ Pus)
Tests	Microscopy & Culture
Sample type	• Sputum
	• BAL
	• Tissue
	• Pus
Collection Container	Universal (60ml wide-mouth, metal top)
Sample collection	 Sputum specimens should be collected early morning. Specimens should be sent to the laboratory without delay during normal working hours. Outside of normal working hours specimens should be refrigerated.
Turnaround Time	2 days for microscopy
	6 weeks for culture
Limitations	N/A
Out-of-Hours-Testing	No
Additional Comments	NOTE: DO NOT send samples in formalin
	TB culture is performed at University Hospital Southampton.

Urine Culture

Investigation Urine Culture

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Document Title: Microbiology User Handbook Q-Pulse Index: MIC-POL-15 Revision Number: 2.1 Author: Olutove, Yinka



Revision Number: 2.1	Author: Olutoye, Yinka
Tests	Microscopy & Culture
Sample type	Urine
Collection Container	Green top tube with boric acid.
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	Container without boric acid
	Urine z
	Universal (white top)
Sample collection	No special requirements.
	• Specimens should be sent to the laboratory without delay during
	normal working hours.
	Outside of normal working hours samples should be
	refrigerated.
Turnaround Time	4 days
Limitations	Delays in transportation may affect the recovery of pathogens.
Out-of-Hours-Testing	No
Additional Comments	Please state in clinical details:
	• Sample type: MSU/ CSU/ SPA/ Bag/ Ileal conduit sample.
	 Antibiotic use (recent and/or intended): helps with
	interpretation of results and guides further work up.
	 For more detail guidance, please refer to:
	https://www.gov.uk/government/publications/urinary-tract-infection-
	<u>uti-diagnosis</u>

Urinary Parasitology (Schistosomiasis)

Investigation	Microscopy for Schistosoma haematobium
Tests	Microscopy
Sample type	Urine
Collection Container	Universal (white top)
Sample collection	 Please collect the sample around 10AM – 2PM after 15 minutes of light exercise.
	 A minimum volume of 10mL is required.
Turnaround Time	5 days
Limitations	In urinary schistosomiasis, very few ova are present in the urine. The number of ova in the urine varies throughout the day, being highest in urine obtained between 10am and 2pm. In patients with haematuria, eggs may be found trapped in the blood and mucus in the terminal portion of the urine specimen. It is therefore preferable to obtain total urine collected over the time period between 10am and 2pm. Alternatively, a 24hr collection of terminal samples of urine may be helpful.
Out-of-Hours-Testing	No
Additional Comments	Sterile containers without boric acid must be used

Active Date: 13/06/2025


Document Title: Microbiology User Handbook Q-Pulse Index: MIC-POL-15 **Revision Number: 2.1** Author: Olutoye, Yinka **13.2 Antigen and Molecular tests**

Investigation	16s PCP PCR/18s Pan-fungal PCR
Tests	PCR
Sample type	Any (including fixed tissue).
	Preferably from normally sterile sites.
Collection Container	Universal (white top)
Sample collection	 Unless stated otherwise in the test table, please send at least 200µL of liquid sample for testing, preferably 500µL to facilitate additional extraction and testing. Low volume samples may be diluted and tested, but reports will bear a caveat regarding the potential effect on assay sensitivity. For tissue samples, please send a matchstick head sized piece of the appropriate tissue in a sterile container.
Turnaround Time	10-14 days
Limitations	Sending sufficient sample is imperative when requesting multiple tests.
Out-of-Hours-Testing	No
Additional Comments	This test is performed at Micropathology University of Warwick

Acanthamoeba Detection

Investigation	Acanthamoeba detection
Tests	Culture and PCR
Sample type	Corneal scrape/biopsy
	Corneal swabs
	Corneal fluids
	 Contact lenses (to be sent in the lens case).
Collection Container	Sterile plastic bijoux container
Sample collection	 The material from a corneal scrape/biopsy should be collected with a needle or blade and rinsed into a small volume (1-2 mL) of sterile saline/distilled water in a small (<5mL) sterile vial/tube. Remove blades or needles as soon as possible after rinsing and before sending. For corneal swabs, do not send dry swabs, please add a small.
	 For corneal swabs, do not send dry swabs, please add a small volume (1-2 mL) of sterile saline or sterile distilled water to the swab to prevent drying.
Turnaround Time	10-14 days
Limitations	 Reference laboratory can also perform culture from contact lenses or fluids; isolation from these specimens, whilst suggestive, does not necessarily implicate the amoeba as causing the patient's symptoms.

Active Date: 13/06/2025



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Out-of-Hours-Testing	No
Additional Comments	This test is performed at London School of Hygiene and Tropical
	Medicine

Adenovirus PCR

This test is used for the diagnosis of acute disease.

Investigation	Adenovirus PCR
Tests	PCR
Sample type	DEPENDS ON ADENOVIRUS TARGET TYPE
	EDTA blood sample (quantitative, immunocompromised
	patients)
	Respiratory secretions
	Eye swabs
	CSF (qualitative)
	Faeces
	Broncho-alveolar lavage
Collection Container	Purple top blood tube
	Viral swabs in viral transport medium
	Sterile plastic bijoux container
Sample collection	Send a viral (green top or red top) swab of vesicle fluid or
	affected mucous membranes. Faeces specimen may be passed
	into a clean, dry, disposable bedpan or similar container and
	transferred to an appropriate collection container.
	 Specimens should be sent to the laboratory during normal
	working hours.
	 Outside of normal working hours samples should be
	refrigerated.
Turnaround Time	10-14 days
Limitations	• False negatives may occur for a variety of reasons, for example
	inappropriate timing of sample collection, inappropriate sample,
	presence of organism below the detectable limit of the assay.
	Towards the limit of detection of an assay sampling variation will
	result in lower reproducibility.
	New and emerging variants may also occur which may not be
	detected by this assay.
Out-of-Hours-Testing	No
Additional Comments	This test is performed at UKHSA Bristol.

Chlamydia trachomatis Infection

Investigation	Detection of Chlamydia trachomatis DNA in urine and genital swabs
Tests	PCR

Document Title: Microbiology User Handbook Q-Pulse Index: MIC-POL-15



Revision Number: 2.1	Author: Olutoye, Yinka NHS Foundation
Sample type	Urine
	Endo-cervical swab
	HVS
	Vulvo-vaginal swabs.
	• Eye
	Throat
	Rectum
Collection Container	Cobas PCR urine tube (Yellow Top)
conection container	
	/* +1 l+
	Universal white top
	1 Contraction of the second se
	VE " manual
	Cobas PCR female swab kit (Yellow Top)
	32 111 4
- I II - I	
Sample collection	DO NOT USE CONTAINERS WITH BORIC ACID
	Specimens should be collected and handled following the
	recommended guidelines on the collection kits. Illustrated
	collection procedure is provided on the back of the collection
	kits.
	• Specimens should be sent to the laboratory without delay during
	normal working hours.
Turnaround Time	7 days
Limitations	Clinical details are essential for processing.
Out-of-Hours-Testing	No
Additional Comments	• Ensure that the lid of the Cobas PCR tube is securely tightened,
	to prevent sample from leaking.
	• The laboratory now screens for <i>Neisseria gonorrhoeae</i> for both
	hospital and community patients. If you do NOT wish to have N.
	gonorrhoeae tested on individual patients, please make this
	clear on the request form (in the clinical details box).
	swab and one for the transport media contained in the pack.
	Note that the expiry date of the swab may differ by some
	months to that of the transport media. It is usually the media
	which has the shorter expiry date.
	• On the Cobas PCR chlamydia swab, the expiry date can be found
	at the bottom of the blister pack, below the Lot number on the
	pack. The date is printed in the reverse order to that we
	normally use in the UK, i.e., YEAR/ MONTH, so March 2023
	would appear as 2023/03. Please return any out-of-date swabs
	to the Microbiology Laboratory and request replacements as
	required.
	NOTE: urine testing for chlamydia in women has been known to produce
	false results. Please contact the Microbiology Laboratory to discuss
	before submitting urine samples from women.
ctive Date: 13/06/2025	

Active Date: 13/06/2025

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Author: Olutoye, Yinka

Chlamydia trachomatis (LGV)

Lymphogranuloma venereum (LGV) is a sexually transmitted infection caused by a particular strain of *Chlamydia* bacteria.

Investigation	Chlamydia trachomatis (LGV)
Tests	PCR
Sample type	Rectal swab
	Throat swab
	Vaginal swab
	Penile swab.
Collection Container	Cobas transport media
	23 Mill specca and
Sample collection	No special requirements.
	Specimens should be sent to the laboratory without delay during
	normal working hours.
	 Outside of normal working hours samples should be
	refrigerated.
Turnaround Time	10-14 days
Limitations	Must be Chlamydia trachomatis positive and symptomatic.
Out-of-Hours-Testing	No
Additional Comments	This test is performed at UKHSA Colindale (STIRL)

COVID-19 PCR

Investigation	Covid-19 PCR
Tests	PCR
Sample type	Nose swab
	Throat swab
	Nasopharyngeal swab
Collection Container	Viral swabs in viral transport medium
Sample collection	No special requirements.
	• Specimens should be sent to the laboratory during normal
	working hours.
Turnaround Time	1 Day.
Limitations	
Out-of-Hours-Testing	Yes, performed by POCT team.

Cryptococcus Antigen

Investigation	Cryptococcus Antigen detection
Tests	Antigen detection
Sample type	Clotted blood
	• CSF
	Biopsy.

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Collection Container	Yellow top (minimum volume 3.5 ml)
	Sterile plastic bijoux container
	Sterile universal container
Sample collection	 Specimens should be sent to the laboratory without delay during normal working hours.
	 Outside of normal working hours samples should be refrigerated.
Turnaround Time	10-14 days
Limitations	Only tested on immunocompromised (including HIV positive) patients.
Out-of-Hours-Testing	No
Additional Comments	This test is performed at the UKHSA Bristol

Detection of Viruses in CSF samples

(Standard: HSV, Varicella Zoster, Enterovirus. Children: Parechoviruses and HHV-6)

Investigation	Detection of viruses in CSF samples
Tests	Viral PCR
Sample type	CSF
Collection Container	2 sterile bijoux containers.
	Send 1st and 3rd samples, appropriately labelled.
Sample collection	 Dispense CSF (minimum 0.5ml in each bottle) into single use containers and label in order.
	 Specimens should be sent to the laboratory without delay during normal hours.
	 Outside of normal hours samples should be placed in the
	pathology reception and the on-call Microbiology Biomedical
	Scientist contacted through switchboard.
Turnaround Time	10-14 days
Limitations	
Out-of-Hours-Testing	No
Additional Comments	 Send if suspected viral meningitis.
	• Laboratory may send this sample to reference laboratory if CSF
	cell count and CSF biochemistry suggests likely viral meningitis.

Full Respiratory Panel (PCR)

(Standard: Influenza A, B, RSV and SARS)

Investigation	Detection of respiratory viruses
Tests	PCR
Sample type	Nasopharyngeal swab

Active Date: 13/06/2025

Document Title: Microbiology User Handbook

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Revision Number: 2.1	Author: Olutoye, Yinka NHS Foundation Trust
	Nose and throat swab in VTM
Collection Container	Green topped swab
	Red topped swab
Sample collection	No special requirements.
	• Specimens should be sent to the laboratory without delay during
	normal working hours.
	Outside of normal working hours samples should be
	refrigerated.
Turnaround Time	1-2 days
Limitations	
Out-of-Hours-Testing	No
Additional Comments	Samples are firstly tested on the Cepheid GeneXpert PCR
	platform to rule out SARS, RSV, Flu A and Flu B infections.
	If the sample is negative for these viruses, Consultant
	Microbiologist decides whether it needs testing for full
	respiratory panel on Biofire PCR platform.
	Not tested routinely- requested by Consultant Microbiologist

Meningococcal PCR

Investigation	Meningococcal PCR
Tests	DNA detection
Sample type	Older children/adults: CSF
	Young children: EDTA blood
Collection Container	Sterile plastic bijoux container
	Purple top (minimum volume 4 ml)
	Pink top
	S NAMES AND A STRATT
Sample collection	Specimens should be sent to the laboratory during normal
	working hours.
	Outside of normal working hours samples should be
	refrigerated.
Turnaround Time	10-14 days
	(Positive result will be phoned earlier)
Limitations	
Out-of-Hours-Testing	No
Additional Comments	Requests must be clearly indicated.
	• Where a CSF sample is available, this should be sent in addition
	to an EDTA blood sample.
	 Clinical details are essential for processing.
	This test is performed at UKHSA Manchester (MMMP)

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Review Date: 13/06/2027

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Author: Olutoye, Yinka

Monkeypox (Mpox) PCR

In the first instance, suspected cases must be discussed with Consultant Microbiologist. Make sure that the request form indicates the risk factors and the reason that Mpox is suspected.

Investigation	Monkeypox PCR
Tests	PCR
Sample type	 Viral swab EDTA Throat swab in VTM Urine.
Collection Container	Green topped swab Red topped swab Purple top (minimum volume 4 ml) Universal (White top)
Sample collection	 Collect specimens as per normal procedure. Specimens should be double bagged, with the request form in the outer bag. Specimens should be sent to the laboratory without delay.
Turnaround Time	10-14 days
Limitations	
Out-of-Hours-Testing	No.
Additional Comments	 Suspected cases need to be discussed with the Consultant Microbiologist first. This test is performed at UKHSA Colindale (RIPL).

Mycobacterium tuberculosis TB Fast Track

Investigation	M. tuberculosis Fast Track test
Tests	Detection of <i>M. tuberculosis</i> complex by molecular amplification
	techniques
Sample type	CSF (min 0.5mL Whole CSF)
	• Wax blocks (for wax blocks, an annotated diagram or slide is
	required indicating the area where AFB were seen, or testing
	should be performed from)
	Sputum
Collection Container	Sterile plastic bijoux container
	Universal (60ml wide-mouth, metal top)



Sample collection	No special requirements.
	 Specimens should be sent to the laboratory during normal
	working hours.
	 Outside of normal working hours samples should be
	refrigerated.
Turnaround Time	10-14 days
Limitations	
Out-of-Hours-Testing	No
Additional Comments	 To be discussed with consultant Microbiologist before being requested. This test is performed at National Mycobacterium Reference Service (NMRS)

Mycobacterium tuberculosis TB T-SPOT

Investigation	TB (Mycobacterium tuberculosis) T-SPOT
Tests	Gamma interferon test
Sample type	X2 Lithium blood
Collection Container	Green top (minimum volume 5 ml)
Sample collection	Specimens not to be refrigerated. Specimens should be sent to the
	laboratory without delay.
	Sample volume:
	 Patient <2 years old: 2mL
	 Patient 2-10 years old: 4 mL
	 Patient >10 years old: 6 mL
Turnaround Time	24-48 hrs
Limitations	 On agreement by Consultant Microbiologist only.
	 Clinical details are essential for processing.
	Monday to Friday ONLY.
	Please fill a Revvity Reference Laboratory request form.
	• Samples must arrive in lab by 16:00 hrs and have been taken
	that morning.
Out-of-Hours-Testing	No
Additional Comments	• This test is performed at Revvity Reference Laboratory, Oxford.

Mycoplasma genitalium PCR

Investigation	Detection of Mycoplasma genitalium rRNA
Tests	PCR
Sample type	Swab in VTM media
	Cobas media
	• Urine (in green tube or sterile universal container).
Collection Container	Green topped swab

Active Date: 13/06/2025

Document Title: Microbiology User Handbook Q-Pulse Index: MIC-POL-15 Revision Number: 2.1 Author: Olutoye, Yinka

Red topped swab IND LADI 42 Cobas transport media (swab) Barrown M Ba Cobas transport media (for urine) - + + | |+ Universal (White top) Sample collection No special requirements. • Specimens should be sent to the laboratory during normal • working hours. Outside of normal working hours samples should be refrigerated. Turnaround Time 10-14 days Limitations **Out-of-Hours-Testing** No Additional Comments This test is performed at UKHSA Colindale (BRD)

NHS Foundation Trust

Parasitology investigation for Enterobius vermicularis

Investigation	Microscopy for Enterobius vermicularis
Tests	Microscopy
Sample type	Sellotape slide
	Perianal swab.
Collection Container	Laboratory no longer supplies collection kits.
	Sellotape slide
	 Cotton-wool swab in dry container.
Sample collection	Sellotape slide
	Apply clear Sellotape to the perianal region, pressing the adhesive side
	of the tape firmly against the left and right perianal folds several times;
	the tape can be wrapped around a tongue depressor to aid specimen
	collection. Smooth the tape back on the slide, adhesive side down.
	Perianal swab
	Perianal specimens are best obtained in the morning before bathing or
	defecation. Three specimens should be taken on consecutive days
	before pinworm infection is ruled out. Cotton-wool swab in dry
	container should be used for collection.
	Spread buttocks apart and rub the moistened cotton wool swab over the
	area around the anus, but do not insert into the anus. Place cotton wool
	swab back in its container (no transport medium required).
	Occasionally, an adult worm may be collected from a patient and sent in
	saline or water for identification.

Active Date: 13/06/2025 Review Date: 13/06/2027 CONTROLLED DOCUMENT; UNCONTROLLED WHEN PRINTED OR COPIED



	Specimens should be sent to the laboratory without delay during normal working hours. Outside of normal working hours samples should be refrigerated.
Turnaround Time	6 days
Limitations	
Out-of-Hours-Testing	No
Additional Comments	

RSV Detection

Investigation	RSV detection
Tests	PCR
Sample type	Nasopharyngeal aspirate
	• Viral swab.
Collection Container	Trap bottle
	Green topped swab
	Red topped swab
Sample collection	No special requirements.
	 Specimens should be sent to the laboratory during normal
	working hours.
	 Outside of normal working hours samples should be
	refrigerated.
Turnaround Time	1 day
Limitations	Clinical details are essential for processing
Out-of-Hours-Testing	No
Additional Comments	

Urine Antigen Tests: Pneumococcal and Legionella

Investigation	Detection of Legionella pneumophila serogroup 1 and Streptococcus
	pneumoniae antigen in urine
Tests	Antigen detection
Sample type	Urine
Collection Container	Universal (white top) or
	yellow top urine collection tubes
	Unitie Z
	Green top tube with boric acid.

Active Date: 13/06/2025

Review Date: 13/06/2027

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Document Title: Microbiology User Handbook Q-Pulse Index: MIC-POL-15



13.3 Serology and Confirmatory PCR tests

Antenatal (booking blood) serology	
Investigation	 Hepatitis B surface antigen (qualitative)
	 HIV-1 and 2 antibodies and HIV p24 antigen (qualitative)
	Treponema pallidum antibody (qualitative)
Tests	Antibody/ Antigen detection
Sample type	Clotted blood
Collection Container	Yellow top (minimum volume 3.5 ml)
Sample collection	No special requirements.
	• Specimens should be sent to the laboratory without delay during
	normal working hours.
	 Outside of normal working hours samples should be
	refrigerated.
Turnaround Time	5 days
Limitations	Haemolysis may affect result.
Out-of-Hours-Testing	No
Additional Comments	 Indicate clearly ALL tests required.
	 Indicate clearly in the clinical details that sample is antenatal
	screening or booking blood.
	 Indicate if patient is a 'late booker'.

Antenatal (booking blood) Serology

Anti-streptolysin titre (ASO Titre)

This test is used to determine past or current infection.

Investigation	Anti-streptolysin titre (ASO Titre)
Tests	Toxin Antibody detection
Sample type	Clotted blood
Collection Container	Yellow top (minimum volume 3.5 ml)
Sample collection	No special requirements.
	 Specimens should be sent to the laboratory without delay during normal working hours.
	 Outside of normal working hours samples should be
	refrigerated.
Turnaround Time	7 days
Limitations	Clinical details are essential for processing.

Active Date: 13/06/2025



Revision Number: 2.1	Author: Olutoye, Yinka
	 Haemolysis can significantly affect the result.
Out-of-Hours-Testing	No
Additional Comments	None

Avian Precipitins

This test is used to determine past or current infection.

Investigation	Avian antibodies (precipitins)
Tests	Antibody detection
Sample type	Clotted blood
Collection Container	Yellow top (minimum volume 3.5 ml)
Sample collection	No special requirements.
	• Specimens should be sent to the laboratory without delay during
	normal working hours.
	 Outside of normal working hours samples should be
	refrigerated.
Turnaround Time	10-14 days
Limitations	Haemolysis may affect result.
Out-of-Hours-Testing	No
Additional Comments	This test is performed at The Royal Wolverhampton NHS Trust (BCPS).

Beta-glucan

This test is used for the diagnosis of fungal infection and exclusion of fungal infection (if negative).

	Rota glucan antigon detection
Investigation	Beta-glucan antigen detection
Tests	Determination of the presence of 1-3 Beta- D- glucan (fungal cell wall
	antigen) in serum
Sample type	Clotted blood
Collection Container	Yellow top (minimum volume 3.5 ml)
Sample collection	No special requirements.
	• Specimens should be sent to the laboratory without delay during
	normal working hours.
	Outside of normal working hours samples should be
	refrigerated.
Turnaround Time	10-14 days
Limitations	
Out-of-Hours-Testing	No
Additional Comments	• Sample must be <48 hrs by the time it reaches the reference
	laboratory and must be a primary sample.
	, , , ,
	 Please note, a BAL sample is not suitable
	This test is performed at UKHSA Bristol.

BK Quantitative PCR virus (Renal patients)

This test is used for the diagnosis of acute disease.

Investigation	BK quantitative PCR
Tests	PCR (quantitative)

Active Date: 13/06/2025

Document Title: Microbiology User Handbook Q-Pulse Index: MIC-POL-15

	NHS Foundation
Revision Number: 2.1	Author: Olutoye, Yinka
Sample type	X 2 EDTA blood samples
Collection Container	Purple top (minimum volume 500µl)
Sample collection	No special requirements.
	 Specimens should be sent to the laboratory without delay during normal working hours.
	 Outside of normal working hours samples should be refrigerated.
Turnaround Time	10-14 days
Limitations	
Out-of-Hours-Testing	No
Additional Comments	• Clinical details must state that it is a renal screening, and sample should be sent to QA Hospital Portsmouth.
	• This test is performed at Queen Alexandra Hospital Portsmouth.

BK Quantitative PCR virus (Non-renal patients)

Investigation	BK quantitative PCR
Tests	PCR (quantitative)
Sample type	X 2 EDTA blood samples
Collection Container	Purple top (minimum volume 500µl)
Sample collection	No special requirements
	• Specimens should be sent to the laboratory without delay during
	normal working hours.
	 Outside of normal working hours samples should be
	refrigerated.
Turnaround Time	10-14 days
Limitations	
Out-of-Hours-Testing	No
Additional Comments	For non-renal patients or not specified to go to QA Hospital
	Portsmouth.
	This test is performed at UKHSA Bristol.

Bordetella pertussis serology

-	
Investigation	Bordetella pertussis serology
Tests	Anti-PT IgG antibody detection
Sample type	Clotted blood
Collection Container	Yellow top (minimum volume 3.5 ml)
Sample collection	No special requirements.
	 Specimens should be sent to the laboratory without delay during normal working hours.
	 Outside of normal working hours samples should be refrigerated.

Active Date: 13/06/2025

Document Title: Microbiology User Handbook Q-Pulse Index: MIC-POL-15 **Revision Number: 2.1** Author: Olutoye, Yinka



Turnaround Time	10-14 days
Limitations	 Samples should be taken 2 weeks after onset of paroxysmal coughing. Pernasal swabs are the most reliable way of making the diagnosis of whooping cough.
Out-of-Hours-Testing	No
Additional Comments	 Pertussis serology is usually more useful in adults presenting with a prolonged cough. Please state onset date. Vaccination history may help interpretation of results. This test is performed at UKHSA Bristol.

Bordetella pertussis PCR

Investigation	Bordetella pertussis PCR
Tests	PCR
Sample type	Pernasal charcoal swab
	 Nasopharyngeal aspirate (NPA)
	Throat swab in VTM
Collection Container	Pernasal swab for whooping cough
	Viral swabs in viral transport medium
Sample collection	No special requirement.
	Specimens should be sent to the laboratory without delay during normal
	working hours.
	Outside of normal working hours samples should be refrigerated.
Turnaround Time	10-14 days
Limitations	Please state vaccination history if known. If sample is urgent, check with
	a Consultant Microbiologist if Biofire PCR testing is required.
Out-of-Hours-Testing	No
Additional Comments	This test is performed at Micropathology University of Warwick

Brucella serology

Investigation	Brucella serology
Tests	Antibody detection
Sample type	Clotted blood
	• CSF
Collection Container	Yellow top (minimum volume 3.5 ml)
	Sterile plastic bijoux container
Sample collection	No special requirements.

Active Date: 13/06/2025

Review Date: 13/06/2027

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CMV Avidity

This test is used to aid differentiation of primary and secondary reactivation of CMV.

Investigation	CMV Avidity
Tests	Enzyme-linked immunosorbent assay (ELISA)
Sample type	Clotted blood
Collection Container	Yellow top (minimum volume 3.5 ml)
Sample collection	No special requirements.
	• Specimens should be sent to the laboratory without delay during
	normal working hours.
	 Outside of normal working hours samples should be
	refrigerated.
Turnaround Time	10-14 days
Limitations	
Out-of-Hours-Testing	No
Additional Comments	Must be CMV IgG positive
	This test is performed at UKHSA Bristol.

CMV IgG and/or CMV IgM

These tests are used for the diagnosis of acute/recent or reactivated disease (IgM) or if evidence of past infection/exposure required (IgG).

pase infection, exposure i	
Investigation	CMV IgG and/or CMV IgM
Tests	Antibody detection
Sample type	Clotted blood
Collection Container	Yellow top (minimum volume 3.5 ml)
Sample collection	 No special requirements. Specimens should be sent to the laboratory without delay during normal working hours. Outside of normal working hours samples should be refrigerated.
Turnaround Time	6 days
Limitations	
Out-of-Hours-Testing	No

Active Date: 13/06/2025

Review Date: 13/06/2027

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Additional Comments	Clinical details are essential for processing. Clearly state whether
	screen or suspected infection.
	Please provide sufficient clinical details.

CMV PCR

This test is used for the diagnosis of acute disease. For diagnosis of congenital CMV send neonatal urine sample within first three weeks of life.

unite sample within hist	
Investigation	CMV PCR
Tests	PCR
Sample type	EDTA blood
	CSF
	Eye fluid
	• Sputum
	• BAL
Collection Container	Purple top (minimum volume 500µl)
	Universal (white top) or yellow top (minimum volume 5ml)
Sample collection	No special requirements.
	• Specimens should be sent to the laboratory without delay during
	normal working hours.
	 Outside of normal working hours samples should be
	refrigerated.
Turnaround Time	10-14 days
Limitations	
Out-of-Hours-Testing	No
Additional Comments	Clinical details are essential for processing.
	This test is processed at UKHSA Bristol.

EBV Serology

Investigation	EBV Serology
Tests	Antibody detection
Sample type	Clotted blood
Collection Container	Yellow top (minimum volume 3.5 ml)
Sample collection	No special requirements.
	• Specimens should be sent to the laboratory without delay during
	normal working hours.
	 Outside of normal working hours samples should be
	refrigerated.
Turnaround Time	5 days
Limitations	
Out-of-Hours-Testing	No
Additional Comments	Clinical details are essential for processing

This test is used for the diagnosis of acute disease.

Investigation	EBV PCR
Tests	PCR
Sample type	EDTA blood
Collection Container	Purple top (minimum volume 4 ml)
Sample collection	No special requirements.
	• Specimens should be sent to the laboratory without delay during
	normal working hours.
	 Outside of normal working hours samples should be
	refrigerated.
Turnaround Time	10-14 days
Limitations	 False negatives may occur for a variety of reasons, for example inappropriate timing of sample collection, inappropriate sample, presence of organism below the detectable limit of the assay.
	 Towards the limit of detection of an assay sampling variation will result in lower reproducibility.
	 New and emerging variants may also occur which may not be detected by this assay.
Out-of-Hours-Testing	No
Additional Comments	 Clinical details are essential for processing.
	This test is processed at UKHSA Bristol.

Fertility Screening

Investigation	Fertility screening
Tests	• HIV
	Hepatitis B core antibody
	Hepatitis B Surface antigen
	Hepatitis C antibody.
Sample type	Clotted blood
Collection Container	Yellow top (minimum volume 3.5 ml)
Sample collection	No special requirements.
	 Specimens should be sent to the laboratory during normal working hours.
	 Outside of normal working hours samples should be refrigerated.
Turnaround Time	10-14 days
Limitations	
Out-of-Hours-Testing	No
Additional Comments	This test is processed at UKHSA Bristol.

Functional Antibodies (Pneumococcal IgG, Haemophilus IgG, Tetanus IgG)

Investigation	Functional Antibodies
Tests	Pneumococcal IgG

Active Date: 13/06/2025

Document Title: Microbiology User Handbook Q-Pulse Index: MIC-POL-15



Fungal precipitins

Investigation	Fungal precipitins
Tests	Antibody detection
Sample type	Clotted blood
Collection Container	Yellow top (minimum volume 3.5 ml)
Sample collection	No special requirements.
	Specimens should be sent to the laboratory during normal
	working hours.
	 Outside of normal working hours samples should be
	refrigerated.
Turnaround Time	10-14 days
Limitations	
Out-of-Hours-Testing	No
Additional Comments	Clinical details are essential for processing.
	This test is processed at UKHSA Bristol

Haemophilus influenzae Type B Ab

This test measures the amount of anti-Hib IgG immunoglobulin (antibody) in the blood.

Investigation	Haemophilus influenzae Type B antibodies
Tests	Antibody detection
Sample type	Clotted blood
Collection Container	Yellow top (minimum volume 3.5 ml)
Sample collection	No special requirements.
	 Specimens should be sent to the laboratory during normal working hours.
	 Outside of normal working hours samples should be refrigerated.
Turnaround Time	10-14 days
Limitations	

Active Date: 13/06/2025

Hepatitis A Serology

Hepatitis A IgG test is used to screen for hepatitis past infection or immunity. Positive result indicates exposure at some time.

Hepatitis A IgM test is used for the diagnosis of acute Hepatitis A infection (jaundice in adults).

Investigation	Hepatitis A IgM and IgG
Tests	Antibody detection
Sample type	Clotted blood
Collection Container	Yellow top (minimum volume 3.5 ml)
Sample collection	No special requirements.
	 Specimens should be sent to the laboratory during normal
	working hours.
	 Outside of normal working hours samples should be
	refrigerated.
Turnaround Time	5 days
Limitations	
Out-of-Hours-Testing	No
Additional Comments	Clinical details are essential for processing, especially onset date.

Hepatitis A PCR

• •	Τ
Investigation	Hepatitis A PCR
Tests	PCR
Sample type	Clotted blood
Collection Container	Yellow top (minimum volume 3.5 ml)
Sample collection	No special requirements.
	 Specimens should be sent to the laboratory during normal
	working hours.
	 Outside of normal working hours samples should be
	refrigerated.
Turnaround Time	10-14 days
Limitations	
Out-of-Hours-Testing	No
Additional Comments	• Performed on Hepatitis A positive or equivocal samples.
	• This test is performed at UKHSA Colindale (VRD).

Hepatitis B surface Antibody

This test is used to determine if protective immunity has been achieved following immunisation. Low levels of hepatitis B virus surface antibody may be found in patients who have past infection.

Investigation	Hepatitis B surface antibody detection	
Tests	Antibody detection	
	(For post vaccination)	
Sample type	Clotted blood	

Active Date: 13/06/2025

Document Title: Microbiology User Handbook Q-Pulse Index: MIC-POL-15 Revision Number: 2.1 Author: Olutoye, Yinka

Collection Container Yellow top (minimum volume 3.5 ml)

Salisbury NHS Foundation Trust

Sample collection	 No special requirements. Specimens should be sent to the laboratory during normal
	working hours.
	Outside of normal working hours samples should be
	refrigerated.
Turnaround Time	5 days
Limitations	 Accurate interpretation of this result is reliant upon detailed
	vaccination history and clinical details.
Out-of-Hours-Testing	No
Additional Comments	 Vaccination history is required for full interpretation of result.
	Blood should be tested 6-8 weeks after final dose of Hepatitis B
	vaccination.

Hepatitis B Core Total Antibody

-	
Investigation	Hepatitis B Core Total Antibody detection
Tests	Antibody detection
	(Acute infection/ evidence of natural immunity)
Sample type	Clotted blood
Collection Container	Yellow top (minimum volume 3.5 ml)
Sample collection	No special requirements.
	 Specimens should be sent to the laboratory during normal working hours.
	 Outside of normal working hours samples should be
	refrigerated.
Turnaround Time	5 days
Limitations	
Out-of-Hours-Testing	No
Additional Comments	Clinical details are essential for processing

Hepatitis B Surface Antigen

This test is used for the diagnosis of acute or recent hepatitis B or carrier state.

Investigation	Hepatitis B Surface Antigen detection
Tests	Antigen detection
Sample type	Clotted blood
Collection Container	Yellow top (minimum volume 3.5 ml)
Sample collection	 No special requirements. Specimens should be sent to the laboratory during normal working hours. Outside of normal working hours samples should be refrigerated.
Turnaround Time	5-7 days
Limitations	

Active Date: 13/06/2025

Additional CommentsRequests must be clearly indicated.

Hepatitis B e Antigen and Antibody and Hepatitis B core IgM

These tests are used to assess infective risk level in acute and chronic hepatitis B virus infections.

Investigation	Hepatitis B e Antigen detection	
	Hepatitis B e Antibody detection	
	Hepatitis B core IgM detection	
Tests	Antibody/antigen detection	
Sample type	Clotted blood	
Collection Container	Yellow top (minimum volume 3.5 ml)	
Sample collection	No special requirements.	
	 Specimens should be sent to the laboratory during normal 	
	working hours.	
	 Outside of normal working hours samples should be 	
	refrigerated.	
Turnaround Time	5 days	
Limitations		
Out-of-Hours-Testing	No	
Additional Comments	Requests must be clearly indicated.	
	 Performed on Hepatitis B surface antigen positive and/or 	
	Hepatitis B core total antibody positive.	

Hepatitis B DNA Viral load

luces attend to a	
Investigation	Hepatitis B DNA Viral load
Tests	PCR
Sample type	EDTA blood
Collection Container	Purple top (minimum volume 3 ml)
Sample collection	No special requirements.
	 Specimens should be sent to the laboratory during normal
	working hours.
	 Outside of normal working hours samples should be
	refrigerated.
Turnaround Time	10-14 days
Limitations	
Out-of-Hours-Testing	No
Additional Comments	Requests must be clearly indicated.
	 Performed on Hepatitis B positive samples.
	This test is performed at UKHSA Bristol.

Hepatitis C Antibody

Marker of infection at some time.

Investigation	Hepatitis C Antibody detection
Tests	Detection of Hepatitis C antibody (qualitative).

Active Date: 13/06/2025

Document Title: Microbiology User Handbook Q-Pulse Index: MIC-POL-15 Revision Number: 2.1 Author: Olutove. Yinka



Revision Number. 2.1	
Sample type	Clotted blood
Collection Container	Yellow top (minimum volume 3.5 ml)
Sample collection	No special requirements.
	• Specimens should be sent to the laboratory during normal
	working hours.
	Outside of normal working hours samples should be
	refrigerated.
Turnaround Time	5 days
	Confirmation of positive result: 10-14 days
Limitations	
Out-of-Hours-Testing	No
Additional Comments	Requests must be clearly indicated.
	Confirmation of positive results are sent to UKHSA Bristol.

Hepatitis C PCR Qualitative

Investigation	Hepatitis C PCR Qualitative
Tests	RNA detection by PCR
Sample type	Clotted blood
Collection Container	Yellow top (minimum volume 3.5 ml)
Sample collection	No special requirements.
	 Specimens should be sent to the laboratory during normal working hours.
	 Outside of normal working hours samples should be
	refrigerated.
Turnaround Time	10-14 days
Limitations	
Out-of-Hours-Testing	No
Additional Comments	Requests must be clearly indicated.
	This test is performed at UKHSA Bristol.

Hepatitis C Viral load/Genotype

Investigation	Hepatitis C Genotype
Tests	Genotype detection by PCR
Sample type	2 x EDTA blood
Collection Container	Purple top (minimum volume 4 ml)
Sample collection	 No special requirements. Specimens should be sent to the laboratory during normal working hours. Outside of normal working hours samples should be refrigerated.
Turnaround Time	10-14 days
Limitations	

Active Date: 13/06/2025 Review Date: 13/06/2027 CONTROLLED DOCUMENT; UNCONTROLLED WHEN PRINTED OR COPIED

Document Title: Microbiology User Handbook Q-Pulse Index: MIC-POL-15



		NHS Foundation
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Out-of-Hours-Testing	No	
Additional Comments	 Requests must be clearly indicated. 	
	• Patient must be HCV positive with active infection.	
	• This test is performed at UKHSA Bristol.	

Hepatitis D (Delta agent)

Investigation	Hepatitis D (Delta agent)
Tests	Antibody detection
	• PCR.
Sample type	Clotted sample
Collection Container	Yellow top (minimum volume 3.5 ml)
Sample collection	No special requirements.
	 Specimens should be sent to the laboratory during normal
	working hours.
	 Outside of normal working hours samples should be
	refrigerated.
Turnaround Time	10-14 days
Limitations	
Out-of-Hours-Testing	No
Additional Comments	Request must be clearly indicated.
	• Sample MUST be Hepatitis B positive.
	This test is performed at UKHSA Colindale (VRD)

Hepatitis E IgM and IgG

Investigation	Hepatitis E IgM and IgG
Tests	Antibody detection
	• PCR.
Sample type	Clotted sample
Collection Container	Yellow top (minimum volume 3.5 ml)
Sample collection	No special requirements.
	 Specimens should be sent to the laboratory during normal
	working hours.
	 Outside of normal working hours samples should be
	refrigerated.
Turnaround Time	7-10 days if confirmation is required
Limitations	
Out-of-Hours-Testing	No
Additional Comments	Request must be clearly indicated.
	 ALT needs to be >100Positive samples are sent to UKHSA
	Colindale (VRD) for PCR testing.

Herpes Simplex Virus 1 and 2 PCR (Qualitative)

For diagnosis of acute disease.

Investigation	Herpes Simplex virus 1 and 2 PCR
-	

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Revision Number: 2.1	Author: Olutoye, Yinka
Tests	PCR
Sample type	Viral swab
Collection Container	Green topped swab
	Red topped swab
Sample collection	No special requirements.
	• Specimens should be sent to the laboratory during normal
	working hours.
	Outside of normal working hours samples should be
	refrigerated.
Turnaround Time	10-14 days
Limitations	• False negatives may occur with inappropriate timing of sample
	collection, inappropriate sample, presence of organism below
	the detectable limit of the assay.
Out-of-Hours-Testing	No
Additional Comments	This test is performed at UKHSA Bristol.

HIV 1/2 Ab/Ag

1 1 0	
Investigation	HIV 1/2 Ab/Ag
Tests	HIV-1 and 2 antibodies and HIV p24 antigen detection(qualitative)
Sample type	Clotted blood
Collection Container	Yellow top (minimum volume 3.5 ml)
Sample collection	No special requirements.
	• Specimens should be sent to the laboratory during normal
	working hours.
	Outside of normal working hours samples should be
	refrigerated.
Turnaround Time	4 days (May take longer if confirmation is required).
Limitations	
Out-of-Hours-Testing	No
Additional Comments	Requests for HIV must be clearly indicated, and the request form
	signed.
	• Positive samples are sent to UKHSA Bristol for confirmation.

HIV 1 Pro-Viral DNA

Investigation	HIV Pro-Viral DNA
Tests	DNA detection in Infants <1 year old
Sample type	2 x EDTA blood
Collection Container	Peach Pink top (paediatric EDTA sample tube)
Sample collection	No special requirements.

Active Date: 13/06/2025

Document Title: Microbiology User Handbook Q-Pulse Index: MIC-POL-15 Revision Number: 2.1 Author: Olutove, Yinka



Revision Number: 2.1	Author: Olutoye, Yinka
	 Specimens should be sent to the laboratory during normal working hours. Outside of normal working hours samples should be refrigerated.
Turnaround Time	10 days
Limitations	
Out-of-Hours-Testing	No
Additional Comments	 Requests for HIV must be clearly indicated, and the request form signed. SAMPLE <u>MUST</u> BE SENT <u>WITH</u> EDTA SAMPLE FROM HIV POSITIVE MOTHER. This test is performed at UKHSA Colindale (VRD)

HIV 1 RNA Viral Load

1 1 • 1 •	
Investigation	HIV 1 RNA Viral Load
Tests	RNA detection in adults and children >1 year old
Sample type	3 x EDTA blood
Collection Container	Purple top (minimum volume 4 ml)
Sample collection	No special requirements.
	 Specimens should be sent to the laboratory during normal
	working hours.
Turnaround Time	1-2 days
Limitations	
Out-of-Hours-Testing	No
Additional Comments	• Requests for HIV must be clearly indicated, and the request form
	signed.
	Sample MUST be HIV 1 positive.

HIV 1 Genotypic Resistance Test

Investigation	HIV Genotypic Resistance Test
Tests	HIV resistance to anti-retroviral therapy
Sample type	10ml EDTA blood
Collection Container	Purple top (minimum volume 4 ml)
Sample collection	 No special requirements. Specimens should be sent to the laboratory during normal working hours. Outside of normal working hours samples should be refrigerated.
Turnaround Time	10-14 days
Limitations	Request from GUM clinic ONLY.
Out-of-Hours-Testing	No
Additional Comments	This test is performed at UKHSA Birmingham (AVTS).

Active Date: 13/06/2025

Document Title: Microbiology User Handbook Q-Pulse Index: MIC-POL-15 Revision Number: 2.1 Author: Olutoye, Yinka



HIV-2 Viral Load

Investigation	HIV-2 Viral load and HIV-2 Pro-Viral testing
Tests	PCR
Sample type	• 1 x Whole Blood EDTA (4-5mL)
Collection Container	Purple top (minimum volume 4 ml)
Sample collection	No special requirements.
Turnaround Time	10-14 days
Limitations	
Out-of-Hours-Testing	No.
Additional Comments	This test is performed at UKHSA Birmingham (AVTS)

HTLV-1 & HTLV-2 Ab

Used to determine past or current infection.

•	
Investigation	HTLV-1 and HTLV-2 Ab
Tests	Antibody detection
Sample type	Clotted blood
Collection Container	Yellow top (minimum volume 3.5 ml)
Sample collection	No special requirements.
	 Specimens should be sent to the laboratory during normal
	working hours.
	 Outside of normal working hours samples should be
	refrigerated.
Turnaround Time	10-14 days.
Limitations	
Out-of-Hours-Testing	No.
Additional Comments	This test is performed at UKHSA Bristol.

Human Herpes Virus-6 DNA Quantitative PCR

frumum mer pes virus		
For diagnosis of HHV-6 in	or diagnosis of HHV-6 infection.	
Investigation	Human Herpes Virus-6 DNA Quantitative PCR	
Tests	PCR	
Sample type	Clotted sample	
	EDTA blood	
Collection Container	Yellow top (minimum volume 3.5 ml)	
	Purple top (minimum volume 4 ml)	
Sample collection	No special requirements.	
	 Specimens should be sent to the laboratory during normal working hours. 	
	 Outside of normal working hours samples should be refrigerated. 	
Turnaround Time	10-14 days	

Document Title: Microbiology User Handbook Q-Pulse Index: MIC-POL-15 Povicion Number: 2.1



Revision Number: 2.1	Author: Olutoye, Yinka
Limitations	 False negatives result may arise due to inappropriate timing of sample collection, inappropriate sample, presence of organism below the detectable limit of the assay.
Out-of-Hours-Testing	No
Additional Comments	This test is performed at UKHSA Colindale (VRD).

Human Herpes Virus-7 DNA Quantitative PCR

For diagnosis of HHV-7 infection.	
Investigation	Human Herpes Virus-7 DNA Quantitative PCR
Tests	PCR
Sample type	EDTA blood
	CSF
	• Serum
Collection Container	Purple top (minimum volume 4 ml)
	Yellow top (minimum volume 3.5 ml)
	Sterile plastic bijoux container
Sample collection	No special requirements.
	Specimens should be sent to the laboratory during normal
	working hours.
	Outside of normal working hours samples should be
	refrigerated.
Turnaround Time	10-14 days
Limitations	False negatives may occur due to inappropriate timing of sample
	collection, inappropriate sample, presence of organism below
	the detectable limit of the assay.
Out-of-Hours-Testing	No
Additional Comments	This test is performed at UKHSA Colindale (VRD).

Human Herpes Virus-8 DNA Quantitative PCR

For diagnosis of HHV-8 infection.

Investigation	Human Herpes Virus-8 DNA Quantitative PCR	
Tests	PCR	
Sample type	EDTA blood	
Collection Container	Purple top (minimum volume 4 ml)	
Sample collection	No special requirements.	
	 Specimens should be sent to the laboratory during normal working hours. Outside of normal working hours samples should be 	
	refrigerated.	
Turnaround Time	10-14 days	

Active Date: 13/06/2025

Document Title: Microbiology User Handbook Q-Pulse Index: MIC-POL-15 Paviaion Number 2.1



Revision Number: 2.1	Author: Olutoye, Yinka
Limitations	• False negatives may occur due to inappropriate timing of sample collection, inappropriate sample, presence of organism below the detectable limit of the assay.
Out-of-Hours-Testing	No
Additional Comments	This test is performed at UKHSA Colindale (VRD).

Leptospiral serology IgM/Leptospiral PCR

Investigation	Leptospiral serology IgM
	Leptospiral PCR
Tests	Antibody detection
	• PCR
Sample type	Clotted blood
	EDTA blood
Collection Container	Yellow top (minimum volume 3.5 ml)
	Purple Top (minimum volume 4 ml)
Sample collection	No special requirements.
	 Specimens should be sent to the laboratory during normal
	working hours.
	 Outside of normal working hours samples should be
	refrigerated.
Turnaround Time	10-14 days
Limitations	
Out-of-Hours-Testing	No
Additional Comments	 Requests must be clearly indicated.
	 Please state date of onset, nature of symptoms and exposure
	history are essential for processing.
	This test is sent to UKHSA Colindale (RIPL).

Lyme (Borrelia burgdorferi) IgG and IgM

Investigation	Borrelia burgdorferi IgG and IgM	
Tests	Antibody detection	
Sample type	Clotted blood	
Collection Container	Yellow top (minimum volume 3.5 ml)	
Sample collection	No special requirements.	
	Specimens should be sent to the laboratory without delay during	
	normal working hours.	
	 Outside of normal working hours samples should be 	
	refrigerated.	
Turnaround Time	4 days	
Limitations		
Out-of-Hours-Testing	No	
Additional Comments	Requests must be clearly indicated.	
	• Other samples (e.g., CSF, joint fluid) by arrangement with	
	Consultant Microbiologist only.	

Active Date: 13/06/2025



• Reactive results are sent to UKHSA Colindale (RIPL).

Measles Serology IgM

Investigation	Measles Serology IgM
Tests	Antibody detection
Sample type	Clotted blood
Collection Container	Yellow top (minimum volume 3.5 ml)
Sample collection	No special requirements.
	 Specimens should be sent to the laboratory during normal
	working hours.
	 Outside of normal working hours samples should be
	refrigerated.
Turnaround Time	10-14 days
Limitations	
Out-of-Hours-Testing	No
Additional Comments	This test is performed at UKHSA Bristol.

Measles Serology IgG

0, 0	
Investigation	Measles Serology IgG
Tests	Antibody detection (evidence of immunity).
Sample type	Clotted blood
Collection Container	Yellow top (minimum volume 3.5 ml)
Sample collection	No special requirements.
	• Specimens should be sent to the laboratory during normal
	working hours.
	Outside of normal working hours samples should be
	refrigerated.
Turnaround Time	5 days
Limitations	
Out-of-Hours-Testing	No
Additional Comments	Requests must be clearly indicated.
	• For acute infection contact local Health Protection Unit (HPU) for
	oral swab test kit

Measles PCR

Investigation	Measles PCR
Tests	PCR
Sample type	Oral fluid
	Throat swabs
	NPA
	 CSF (150μl)
	Urine
	• Tissue.
Collection Container	Green topped viral swab

Active Date: 13/06/2025

Document Title: Microbiology User Handbook Q-Pulse Index: MIC-POL-15 Revision Number: 2.1 Author: Olutoye, Yinka



	Red topped viral swab
	PERSONAL PROPERTY OF THE PERSON OF THE PERSO
	Universal (white top)
	Sterile plastic bijoux container
Sample collection	No special requirements.
	 Specimens should be sent to the laboratory during normal working hours.
	 Outside of normal working hours samples should be refrigerated.
Turnaround Time	10-14 days
Limitations	
Out-of-Hours-Testing	No
Additional Comments	 NOTE: Contact the laboratory before sending a tissue.
	This test is performed at UKHSA Bristol.

Mumps Serology IgG

Investigation	Mumps Serology IgG
Tests	Antibody detection
Sample type	Clotted blood
Collection Container	Yellow top (minimum volume 3.5 ml)
Sample collection	No special requirements.
	 Specimens should be sent to the laboratory during normal
	working hours.
	 Outside of normal working hours samples should be
	refrigerated.
Turnaround Time	5 days
Limitations	
Out-of-Hours-Testing	No
Additional Comments	Requests must be clearly indicated.

Mumps Serology IgM

Investigation	Mumps IgM
Investigation	
Tests	Antibody detection
Sample type	Clotted blood
Collection Container	Yellow top (minimum volume 3.5 ml)
Sample collection	No special requirements.

Active Date: 13/06/2025

Document Title: Microbiology User Handbook Q-Pulse Index: MIC-POL-15



Mumps PCR

Investigation	Mumps PCR
Tests	PCR
Sample type	Mouth/throat swab
Collection Container	Green topped swab
	Red topped swab
Sample collection	No special requirements.
	Specimens should be sent to the laboratory during normal
	working hours.
	 Outside of normal working hours samples should be
	refrigerated.
Turnaround Time	10-14 days
Limitations	
Out-of-Hours-Testing	No
Additional Comments	Clinical details are essential.
	Please state vaccination history.
	This test is performed at UKHSA Colindale (VRD).

Non-indigenous mycoses (e.g., Histoplasma) serology

1 • • • • • • • • • •	
Investigation	Non-indigenous mycoses serology
Tests	Antibody detection
Sample type	Clotted blood
Collection Container	Yellow top (minimum volume 3.5 ml)
Sample collection	No special requirements.
	 Specimens should be sent to the laboratory during normal working hours.
	 Outside of normal working hours samples should be refrigerated.
Turnaround Time	10-14 days
Limitations	

Active Date: 13/06/2025

Document Title: Microbiology User Handbook Q-Pulse Index: MIC-POL-15 Revision Number: 2.1

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Sa	lisbury
NHS Found	ation Trust

Revision Number: 2.1	Author: Olutoye, Yinka	NHS Foundation
Out-of-Hours-Testing	No	
Additional Comments	Travel history is essential.	
	• This test is performed at UKHSA Bristol.	

Parasite disease serology

Various including *Schistosoma*, Amoebic (abscess), *Toxocara*, etc.

Investigation	Serology for various parasites including Schistosoma, Amoebas (abscess),	
	<i>Toxocara</i> , etc.	
Tests	Antibody detection	
Sample type	Clotted blood	
Collection Container	Yellow top (minimum volume 3.5 ml)	
Sample collection	No special requirements.	
	 Specimens should be sent to the laboratory during normal 	
	working hours.	
	 Outside of normal working hours samples should be 	
	refrigerated.	
Turnaround Time	10-14 days	
Limitations		
Out-of-Hours-Testing	No	
Additional Comments	Clinical details including countries visited & dates are essential.	
	Contact duty Consultant Microbiologist if required.	
	• This test is performed at Health Services Laboratory.	

Parvovirus B19 IgG and IgM Serology

0	
Investigation	Parvovirus Serology
Tests	Antibody detection
Sample type	Clotted blood
Collection Container	Yellow top (minimum volume 3.5 ml)
Sample collection	No special requirements.
	• Specimens should be sent to the laboratory during normal
	working hours.
	 Outside of normal working hours samples should be
	refrigerated.
Turnaround Time	10-14 days
Limitations	
Out-of-Hours-Testing	No
Additional Comments	Clinical details are essential for processing.
	• This test is performed at UKHSA Bristol.

Parvovirus B19 PCR

Investigation	Parvovirus B19 PCR
Tests	PCR
Sample type	• EDTA
	• Serum.
Active Date: 13/06/202	5 Review Date: 13/06/2027

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Revision Number: 2.1	Author: Olutoye, Yinka
Collection Container	Purple top (minimum volume 4 ml)
	Yellow top (minimum volume 3.5 ml)
Sample collection	No special requirements.
	 Specimens should be sent to the laboratory during normal
	working hours.
	 Outside of normal working hours samples should be
	refrigerated.
Turnaround Time	10-14 days
Limitations	
Out-of-Hours-Testing	No
Additional Comments	This test is performed at UKHSA Bristol.
Limitations Out-of-Hours-Testing	Outside of normal working hours samples should be refrigerated. 10-14 days No

Pneumococcal PCR

Investigation	Pneumococcal PCR
Tests	DNA detection
Sample type	EDTA blood and/or CSF
Collection Container	Purple top for blood (minimum volume 4 ml)
	Sterile plastic bijoux container
Sample collection	 Dispense CSF (minimum 0.5ml in each bottle) into single use containers and label in order.
	• Specimen should be sent to the laboratory during normal hours.
	Outside of normal hours samples should be placed in the
	pathology reception and the on-call Microbiology Biomedical
	Scientist contacted through switchboard.
	 No special requirements for EDTA samples.
Turnaround Time	10-14 days (positive result will be phoned earlier).
Limitations	
Out-of-Hours-Testing	No
Additional Comments	Requests must be clearly indicated.
	This test is performed at UKHSA Manchester (MMMP)

Pneumococcal Serology

Investigation	Streptococcus pneumoniae antibody detection
Tests	Antibody detection
Sample type	Clotted blood
Collection Container	Yellow top (minimum volume 3.5 ml)
Sample collection	No special requirements.
	 Specimens should be sent to the laboratory during normal working hours.
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Active Date: 13/06/2025

Document Title: Microbiology User Handbook Q-Pulse Index: MIC-POL-15 Revision Number: 2.1 Author: Olutove, Yinka



Revision Number: 2.1	Author: Olutoye, Yinka
	 Outside of normal working hours samples should be refrigerated.
Turnaround Time	10-14 days
Limitations	
Out-of-Hours-Testing	No
Additional Comments	This test is performed at UKHSA Manchester (MMMP)

Rabies Serology

Investigation	Rabies antibody detection
Tests	Antibody detection
Sample type	Clotted blood
Collection Container	Yellow top (minimum volume 3.5 ml)
Sample collection	 No special requirements. Specimens should be sent to the laboratory without delay during normal working hours. Outside of normal working hours samples should be refrigerated.
Turnaround Time	10-14 days
Limitations	
Out-of-Hours-Testing	No
Additional Comments	This test is performed at Animal and Plant Health Agency Rabies
	Laboratory, Weybridge

Rubella Serology

Investigation	Rubella Serology IgG/IgM
Tests	Antibody detection
Sample type	Clotted blood
Collection Container	Yellow top (minimum volume 3.5 ml)
Sample collection	No special requirements.
	• Specimens should be sent to the laboratory during normal
	working hours.
	 Outside of normal working hours samples should be
	refrigerated.
Turnaround Time	5 days
Limitations	
Out-of-Hours-Testing	No
Additional Comments	Clinical details are essential for processing.

Schistosoma Serology

Investigation	Schistosoma antibody detection
Tests	Antibody detection
Sample type	Clotted blood

Document Title: Microbiology User Handbook Q-Pulse Index: MIC-POL-15 Povision Number: 2.1

Q-Pulse Index: IVIIC-POL-	15 NHS Foundation T
Revision Number: 2.1	Author: Olutoye, Yinka
Collection Container	Yellow top (minimum volume 3.5 ml)
Sample collection	No specimen requirements.
	 Specimens should be sent to the laboratory during normal working hours.
	 Outside of normal working hours samples should be refrigerated.
Turnaround Time	10-14 days
Limitations	
Out-of-Hours-Testing	No
Additional Comments	Please state travel history or other relevant clinical details.
	• This test is performed at Health Services Laboratory.

Syphilis Serology

Investigation	Syphilis antibody detection.
Tests	Antibody detection
Sample type	Clotted blood
Collection Container	Yellow top (minimum volume 3.5 ml)
Sample collection	No special requirements.
	 Specimens should be sent to the laboratory without delay during normal working hours.
	Outside of normal working hours samples should be
	refrigerated.
Turnaround Time	4 days, longer if the sample is positive and needs sending to the
	reference laboratory for confirmation.
Limitations	
Out-of-Hours-Testing	No
Additional Comments	Clinical details are essential for processing.
	Positive in-house samples are sent to UKHSA Bristol

Syphilis IgM, RPR, TPPA, alternative EIA and Immunoblots

Investigation	Syphilis IgM, RPR, TPPA, alternative EIA and Immunoblots
Tests	Antibody detection
Sample type	Clotted blood
Collection Container	Yellow top (minimum volume 3.5 ml)
Sample collection	No special requirements.
	Specimens should be sent to the laboratory without delay during
	normal working hours.
	 Outside of normal working hours samples should be
	refrigerated.
Turnaround Time	5-7 days (Longer if sample needs to be sent to Reference Laboratory).
Limitations	
Out-of-Hours-Testing	No
Additional Comments	If sample is positive, it is sent to UKHSA Bristol.

Active Date: 13/06/2025





Author: Olutoye, Yinka

Syphilis PCR	
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Investigation	Syphilis PCR
Tests	Detection of Treponema pallidum DNA
Sample type	Swab in viral transport media
Collection Container	Green topped swab
	Red topped swab
Sample collection	No special requirements.
	 Specimens should be sent to the laboratory without delay during normal working hours.
	 Outside of normal working hours samples should be refrigerated.
Turnaround Time	10-14 days
Limitations	
Out-of-Hours-Testing	No
Additional Comments	This test is performed at PHE Colindale (STIRL).

Toxoplasma gondii Serology

	To a subscript of the state st
Investigation	Toxoplasma gondii antibody detection
Tests	Antibody detection
Sample type	Clotted blood
Collection Container	Yellow top (minimum volume 3.5 ml)
Sample collection	No special requirements.
	• Specimens should be sent to the laboratory without delay during
	normal working hours.
	 Outside of normal working hours samples should be
	refrigerated.
Turnaround Time	5 days, longer if confirmation of positive result is required.
Limitations	
Out-of-Hours-Testing	No
Additional Comments	Clinical details are essential for processing.
	• Confirmation of positive results sent to PHW Swansea.

Tropical Disease Serology (Regional Travel Screen)

Various including Dengue, Q fever, Nile, Rickettsia, Zika virus, Viral haemorrhagic fevers, etc.

Investigation	Tropical disease (e.g., Dengue, VHF, etc.) antibody detection
Tests	Antibody detection
Sample type	Clotted blood
Collection Container	Yellow top (minimum volume 3.5 ml)
Sample collection	No special requirements.
	 Specimens should be sent to the laboratory without delay during normal working hours.
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Revision Number. 2.1	Author: Oldtoye, Tilka
	Outside of normal working hours samples should be
	refrigerated.
Turnaround Time	7-14 days
Limitations	Vaccinations & antibiotics given may affect test results.
Out-of-Hours-Testing	No
Additional Comments	Clinical details including countries visited & dates are essential.
	Contact duty Consultant Microbiologist if required.
	• This test is performed at UKHSA Colindale (RIPL).

Varicella zoster Serology (Chickenpox)

This test is used to determine past chickenpox infection (or vaccination). It indicates immunity.

Investigation	Varicella zoster IgG antibody detection.
Tests	Antibody detection
Sample type	Clotted blood
Collection Container	Yellow top (minimum volume 3.5 ml)
Sample collection	No special requirements.
	• Specimens should be sent to the laboratory without delay during
	normal working hours.
	 Outside of normal working hours samples should be
	refrigerated.
Turnaround Time	5 days normal
	1 day (urgent)
Limitations	Please contact Laboratory if urgent processing is required.
	Please state date of contact.
Out-of-Hours-Testing	Saturday/ Sunday/ Bank Holiday morning (by arrangement only).
Additional Comments	When requesting Varicella zoster antibody following contact
	with chickenpox in both pregnant women and
	immunocompromised patients it is essential that the date the
	patient was in contact with the chickenpox case is stated in the
	clinical details as well as the onset date of the chickenpox case's
	rash as these are used to assess the value of Varicella Zoster
	Immunoglobulin (VZIG) in every case.
	Please contact the laboratory in such cases so that the samples
	can be tested urgently on arrival. This is particularly important
	on Fridays, weekends, and Bank Holidays when staffing is
	reduced. Please include contact number (bleep or extension) so
	the result could be telephoned through to the clinician.
	 For non-immune contacts, VZIG is only available if the result is
	known less than 10 days after contact, otherwise other
	therapeutic options may be required.

Varicella zoster PCR

Investigation	Detection of Varicella zoster DNA
Tests	PCR
Sample type	Viral swab
	• CSF
	Whole EDTA

Document Title: Microbiology User Handbook Q-Pulse Index: MIC-POL-15

Q-Pulse Index: MIC-POL-	15 Salisburg NHS Foundation Trus
Revision Number: 2.1	Author: Olutoye, Yinka
Collection Container	Green topped swab Red topped swab Sterile glass bijoux container Purple top (minimum volume 4 ml)
Sample collection	 For screening, swab should be taken from the affected site. Specimens should be sent to the laboratory without delay during normal working hours. Outside of normal working hours samples should be refrigerated.
Turnaround Time	10-14 days.
Limitations	
Out-of-Hours-Testing	No
Additional Comments	 Please provide sufficient and relevant clinical details (e.g., vesicular rash, shingles). This test is performed at UKHSA Bristol.

13.4 Antibiotic assays

Gentamicin Levels

Gentalincin Levels	
Investigation	Gentamicin Level
Tests	Antibiotic assay
Sample type	Clotted blood
Collection Container	Yellow top (minimum volume 3.5 ml)
Sample collection	No special requirements.
	 Specimens should be sent to the laboratory without delay during normal working hours.
	 Outside of normal working hours samples should be
	refrigerated.
Turnaround Time	1 day
Limitations	
Out-of-Hours-Testing	Yes (must be arranged with on call biomedical scientist in Laboratory
	Medicine).
Additional Comments	• Timing of sample, and drug dose and timing regimen essential
	for interpretation of result
	Refer to gentamicin guidelines on Eolas
	 Please use a green biochemistry request form.
	This test is performed in Laboratory Medicine at Salisbury NHSFT

Active Date: 13/06/2025

Review Date: 13/06/2027

Salisbury



Author: Olutoye, Yinka

Tobramycin Levels

Investigation	Tobramycin Levels
Tests	Antibiotic assay
Sample type	Clotted blood
Collection Container	Yellow top (minimum volume 3.5 ml)
Sample collection	No special requirements.
	 Specimens should be sent to the laboratory without delay during normal working hours.
	 Outside of normal working hours samples should be refrigerated.
Turnaround Time	2-3 days for verbal result, 7 – 10 days for electronic report.
Limitations	
Out-of-Hours-Testing	No
Additional Comments	 Timing of sample, and drug dose and timing regimen essential for interpretation of result.
	 Please make sure that you complete the request form fully and provide dosing details.
	 Please use a black microbiology request form
	 This test is performed at UKHSA Bristol Antimicrobial Reference Laboratory

Amikacin Levels

Investigation	Amikacin Levels
Tests	Antibiotic assay
Sample type	Clotted blood
Collection Container	Yellow top (minimum volume 3.5 ml)
Sample collection	No special requirements.
	 Specimens should be sent to the laboratory without delay during normal working hours
	normal working hours.
	 Outside of normal working hours samples should be refrigerented.
	refrigerated.
Turnaround Time	2.2 days for verbal result
Turnaround Time	2-3 days for verbal result
-	7 – 10 days for electronic report.
Limitations	
Out-of-Hours-Testing	No
Additional Comments	• Timing of sample, and drug dose and timing regimen essential
	for interpretation of result.
	Please make sure that you complete the request form fully and
	provide dosing details.
	 Please use a black microbiology request form.
	This test is performed at UKHSA Bristol Antimicrobial Reference
	Laboratory
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Document Title: Microbiology User Handbook Q-Pulse Index: MIC-POL-15 **Revision Number: 2.1** Author: Olutoye, Yinka



Vancomycin Levels

Pre dose only unless requested by Microbiologist)	
Investigation	Vancomycin Levels
Tests	Antibiotic assay
Sample type	Clotted blood
Collection Container	Yellow top (minimum volume 3.5 ml)
Sample collection	No special requirements.
	• Specimens should be sent to the laboratory without delay during
	normal working hours.
	 Outside of normal working hours samples should be
	refrigerated.
Turnaround Time	1 day
Limitations	
Out-of-Hours-Testing	Yes - during daytime at weekends.
	(Must be arranged with on call biomedical scientist in Laboratory
	Medicine).
Additional Comments	Timing of sample, and drug dose and timing regimen essential
	for interpretation of result.
	 Refer to vancomycin guidelines on Eolas.
	 Please use a green biochemistry request form
	• This test is performed in Laboratory Medicine at Salisbury NHSFT

Teicoplanin level

(Pre dose only as advised by Microbiologist)

(FIE dose only as advised	by wherebiologist)
Investigation	Teicoplanin level
Tests	Antibiotic assay
Sample type	Clotted blood
Collection Container	Yellow top (minimum volume 3.5 ml)
Sample collection	No special requirements.
	Specimens should be sent to the laboratory without delay during
	normal working hours.
	 Outside of normal working hours samples should be
	refrigerated.
Turnaround Time	2-3 days for verbal result, 7 – 10 days for electronic report
Limitations	
Out-of-Hours-Testing	No – unless agreed before the weekend with Consultant Microbiologist
Additional Comments	• Timing of sample, and drug dose and timing regimen essential
	for interpretation of result.
	 Please make sure that you complete the request form fully and
	provide dosing details.
	 Please use a black microbiology request form.
	• This test is performed at UKHSA Bristol Antimicrobial Reference
	Laboratory.

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Other antibiotic level, e.g., Co-trimoxazole

Investigation	Other antibiotic level, e.g., Co-trimoxazole
Tests	Antibiotic assay
Sample type	Clotted blood
Collection Container	Yellow top (minimum volume 3.5 ml)
Sample collection	No special requirements.
	• Specimens should be sent to the laboratory without delay during
	normal working hours.
	Outside of normal working hours samples should be
	refrigerated.
Turnaround Time	2-3 days for verbal result, 7 – 10 days for electronic report
Limitations	
Out-of-Hours-Testing	No
Additional Comments	• Timing of sample, and drug dose and timing regimen essential
	for interpretation of result
	 Pre-arrangement with Consultant Microbiologist ONLY.
	 Please make sure that you complete the request form fully and
	provide dosing details.
	 This test is performed at UKHSA Bristol Antimicrobial Reference Laboratory.

Anti-Fungal drug level

Tiller Tullgar al ag leve	T
Investigation	Anti-fungal drug level
Tests	Anti-fungal assay
Sample type	Red top serum tube (Non-separator)
Collection Container	Red top
	A & Balance
Sample collection	No special requirements.
	• Specimens should be sent to the laboratory without delay during
	normal working hours.
	 Outside of normal working hours samples should be
	refrigerated.
Turnaround Time	2-3 days for verbal result, 7 – 10 days for electronic report.
Limitations	
Out-of-Hours-Testing	No
Additional Comments	• Timing of sample, and drug dose and timing regimen essential for interpretation of result.
	 Pre-arrangement with Consultant Microbiologist ONLY.
	 Please make sure that you complete the request form fully and provide dosing details.
	 Please use a black microbiology request form.
	These tests are done at UKHSA Bristol Antimicrobial Reference Laboratory.



13.5 Family Planning

Sub-fertility semen analysis

Semen analysis is the microscopic examination of the semen to see how many sperm cells there are and whether they function correctly.

Investigation	Sub-fertility semen (Andrology)
Tests	Microscopy (analysis of cells and cell count)
Sample type	Semen sample
Collection Container	Universal (Non-Toxic specimen container - contact laboratory)
Sample collection	• The sample needs to be collected in the morning. Patient must
	abstain from sexual activity for 48 hours (but no longer than 7
	days) before collecting the sample.
	 Sample should be collected into the provided container (it can be callected from CD or fortility and ciclist)
	be collected from GP or fertility specialist).
	 Sample should be collected by masturbation and the entire
	specimen should be collected into the container.
	DO NOT use a sheath or condom for collection as they are
	harmful to sperm.
	• After the collection, please keep the sample close to the body
	whilst travelling as sperm is sensitive to temperature changes.
	• To ensure the test results will be accurate, aim to deliver the
	sample within 1 hour of its production.
Turnaround Time	7 days
Limitations	Samples by appointment only (patient to contact laboratory).
	Fresh sample taken on day of submission.
Out of House Testing	To arrive within 1 hour of being taken.
Out-of-Hours-Testing	No
Additional Comments	Please note: Patient leaflets with instructions on how to take samples for
	Sub-fertility (Semen analysis) samples are available on the Salisbury NHS Foundation Trust MICROGUIDE web site at:
	Diagnostic Semen Analysis (Male Fertility Testing) (Eolas)
	The department runs a weekly andrology clinic on Wednesday mornings.
	There are 6 appointment slots available per week except for days where
	bank holidays occur. Please ring the laboratory on extension 4099 or
	4105 to make an appointment prior to sample collection. Patients
	providing semen samples for Fertility assessment attend with their
	samples and complete a questionnaire to ensure the Andrology service
	complies with UKAS quality requirements.
	Please notify the laboratory if you are unable to attend your
	appointment so that the appointment slot can be offered to other
	patients where possible.
	Please ensure that patients attending for Fertility tests are provided with
	a completed black Microbiology form PLUS a suitable non-toxic wide-
	mouthed sterile container to permit the complete semen sample to be
	captured by the patient. The laboratory provides assembled "collection
	packs" for Fertility patients which are available at all surgeries/ clinics. If
	captured by the patient. The laboratory provides assembled "collection



replacement packs are required, please ring (01722) 429105 to request
replacements.
We advise that the requesting clinician goes through the process with
the patient at the time the form and container are supplied to ensure
the patient understands when and how to collect the sample. This will
help to ensure complete semen sample collection and therefore improve
the accuracy of the test.
Samples received in alternative containers to the issued sterile non-toxin
containers will NOT be processed.
Fertility is a multi-factorial state and it is advised that the semen test
result should be read whilst taking into account other physical and
physiological factors affecting a couple's fertility.

Post vasectomy semen analysis

Post-vasectomy semen analysis is the test done by the laboratory to confirm that "No sperms are seen", however, this test does NOT exclude presence of a very small number of sperms in the sample provided.

Investigation	Post vasectomy semen analysis
Test	Microscopy
Sample type	Semen sample
Collection Container	Universal (Non-Toxic specimen container-contact laboratory)
Sample collection	 The sample needs to be collected in the morning. Patient must abstain from sexual activity for 48 hours (but no longer than 7 days) before collecting the sample. Sample should be collected into the provided container (it can be collected from GP). Sample should be collected by masturbation and the entire specimen should be collected into the container. DO NOT use a sheath or condom for collection as they are harmful to sperm. After the collection, please keep the sample close to the body whilst travelling.
Turnaround Time	3-4 days
Limitations	Fresh sample taken on day of submission. To arrive in Lab between 09:00 and 12:00
Out-of-Hours Testing	No
Additional Comments	Post vasectomy samples can by submitted any weekday (Monday – Friday) except bank holidays. Patients are asked to bring their samples to the pathology reception desk on level 3. No appointment is required. Please note: Patient leaflets with instructions on how to take samples for post-vasectomy samples are available on the Salisbury NHS Foundation Trust MICROGUIDE web site at: <u>post-vasectomy-for-printing-april-2020.docx (live.com)</u> Please ensure that patients attending for post-vasectomy testing are provided with a completed black Microbiology form PLUS a suitable non- toxic wide-mouthed sterile container to permit the complete semen sample to be captured by the patient. We advise that the requesting clinician goes through the process with the patient at the time the form and container are supplied to ensure the patient understands when and how to collect the sample. This will help to
	patient understands when and now to collect the sample. This will help to

Active Date: 13/06/2025



Author: Olutoye, Yinka

ensure complete semen sample collection and therefore improve the accuracy of the test.

14. Point of Care Testing (POCT)

Our Microbiology Department offers Point of Care Testing from 8PM to 6AM every night. POCT team performs SARS-CoV-2, Flu and RSV testing. They are located on level 3, opposite the Accident and Emergency Department.

POCT team uses the Xpert Xpress SARS-CoV-2/Flu/RSV test, which is an automated *in vitro* diagnostic test for the qualitative detection and differentiation of RNA from Flu A, Flu B, RSV and SARS-CoV-2 virus. The assay is performed on the GeneXpert instrument system which performs sample preparation, nucleic acid extraction, amplification, and detection of target sequences in simple or complex samples using real time polymerase chain reaction (RT-PCR) assays.

The assay is suitable for detection of viruses from nasopharyngeal swabs, nasal swabs, or nasal wash/aspirate specimens.

All negative results are phoned to wards and positive results are phoned to the Site Manager.

15. Patient Consent Disclosure

15.1 Laboratory policy on protection of personal information

The Microbiology Department regards the lawful and correct treatment of patients' personal information as vital to maintaining the confidence of users of the service. Our policy is that we will adhere to the principles of data protection described in the Data Protection Act 1998 part 1, 3 (9). As part of the Salisbury District Hospital NHS Foundation Trust, we also work to its governance and data protection policies.

15.2 Patient consent

All samples need to have patients consent, therefore all requests for investigations must include the requesting physician's signature on the request form. All unsigned forms may be returned to the requestor before testing is commenced.

The laboratory does not seek to confirm that informed consent has been obtained for any specimen that is sent for analysis. It is the responsibility of referring clinicians to ensure appropriate consent has been obtained. Requesting a specific test implies patient consent has been obtained. Where this is impossible, testing should only take place when it is in the best interests of the patient. The General Medical Council provides guidance which should be consulted on this issue.

15.3 Medico-legal samples

Any specimen submitted for medico-legal purposes should have documentation accompanying these specimens to provide an unbroken chain of evidence.

Document Title: Microbiology User Handbook Q-Pulse Index: MIC-POL-15 Revision Number: 2.1



Author: Olutoye, Yinka

For further guidance on handling medicolegal samples and preserving the chain of evidence please refer to this website: New guidance for handling medicolegal samples and preserving the chain of evidence (rcpath.org)

15.4 The Human Tissue Act

Salisbury District Hospital NHS Foundation Trust are licensed by the Human Tissue Act (HTA) to undertake examinations of post-mortem samples submitted by clinical consultants and pathologists. Under the license, the samples may be retained until the examination has been completed and in line with the sample retention policies. It is the obligation of the requesting clinician or pathologist to ensure that examination of samples they submit have been requested by the coroner or appropriate consent has been obtained from the deceased person or their relatives. Only the specific examinations requested by the sending clinician or pathologist may be performed.

If additional work on samples from the deceased is thought necessary by the medical microbiologist or virologist, they must obtain written confirmation of consent from the sending departments. All relevant material is stored securely and under conditions which maintain the integrity of the sample if possible and confidentiality is maintained in compliance with Caldicott principles, as are all samples received. Following processing, relevant material is only retained for the period of time specified by the retention policy.

For further information please refer to Human Tissue Act 2004: Human Tissue Act 2004 (legislation.gov.uk)

16 Feedback on our microbiology service and complaints procedure

The laboratory is committed to providing a high-quality service to all service users; however, it understands that aspects of the service may not always meet the requirements of the customer. Should this occur and there be a requirement to give feedback or make a complaint to the laboratory please submit your feedback or complaint in writing to the following:

Lee Phillips	lee.phillips@nhs.net
Head of Pathology	
Joanne Harris	joanne.harris7@nhs.net
Microbiology Laboratory Manager	
Patient Advice Liaison Services (PALS)	<u>sft.pals@nhs.net</u>

17 Associated Documents

- Trust Specimen Transportation Policy
- Infection control policy
- Pneumatic Air Tube Guidance, Salisbury NHS Foundation Trust, Version 2.0, 2024.



Appendix 1- HSE Safety Notice

Risk of exposure of laboratory staff to biological agents due to missing clinical information

Workers at diagnostic laboratories are being put at risk of infection because of missing information on specimen request forms. If specimen request forms do not provide sufficient clinical information, then lab staff cannot identify the appropriate safety measures they need to apply to control exposure and possible infection.

Anyone who completes a specimen request form, like clinicians, nurses, GPs or vets, should include all information that can help lab staff assess potential risks.

The following information is needed to identify risks:

- recent travel history
- consumption of unpasteurised milk or cheese products
- contact with imported animals (in particular dogs from Eastern Europe)
- association with an outbreak scenario
- other relevant clinical symptoms or information

Ensure that specimen request forms contain all relevant clinical information.

<u>Read the safety notice on the risk of exposure to biological agents for full</u> <u>details</u>